

# Package ‘macroSyntR’

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**Type** Package

**Title** Draw Ordered Oxford Grids and Chord Diagrams

**Version** 0.3.3

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**Depends** R (>= 4.1.0)

**Imports** stats, utils, ggplot2, igraph, tidyr, reshape2, dplyr,  
stringr, rlang

**Description** Use standard genomics file format (BED) and a table of orthologs to illustrate synteny conservation at the genome-wide scale. Significantly conserved linkage groups are identified as described in Simakov et al. (2020) <[doi:10.1038/s41559-020-1156-z](https://doi.org/10.1038/s41559-020-1156-z)> and displayed on an Oxford Grid (Edwards (1991) <[doi:10.1111/j.1469-1809.1991.tb00394.x](https://doi.org/10.1111/j.1469-1809.1991.tb00394.x)>) or a chord diagram as in Simakov et al. (2022) <[doi:10.1126/sciadv.abi5884](https://doi.org/10.1126/sciadv.abi5884)>. The package provides a function that uses a network-based greedy algorithm to find communities (Clauset et al. (2004) <[doi:10.1103/PhysRevE.70.066111](https://doi.org/10.1103/PhysRevE.70.066111)>) and so automatically order the chromosomes on the plot to improve interpretability.

**Encoding** UTF-8

**License** GPL-3

**URL** <https://github.com/SamiLh11/macroSyntR>

**BugReports** <https://github.com/SamiLh11/macroSyntR/issues>

**RoxygenNote** 7.2.1

**Suggests** knitr, rmarkdown

**VignetteBuilder** knitr

**NeedsCompilation** no

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compute\_linkage\_groups

*Compute Linkage groups*

---

### Description

This is a function to compute the conserved linkage groups shared between two or more species. It computes the significant associations between chromosomes of all species versus all (pairwise) using the fischer test implemented in compute\_macrosyteny(). It outputs a dataframe shaped as following : sp1.Chr,sp2.Chr,..., spN.chr,n,LGs where n is the number of shared orthologs in the group and LGs are the IDs for the linkage groups

### Usage

```
compute_linkage_groups(orthologs_df)
```

### Arguments

orthologs\_df     dataframe. orthologs with genomic coordinates loaded with load\_orthologs()

### Value

A dataframe object

### Examples

```
# basic usage of compute_linkage_groups:

orthologs_table <- system.file("extdata", "my_orthologs.tab", package="macrosyntR")

my_orthologs <- read.table(orthologs_table, header=TRUE)

my_macrosyteny <- compute_linkage_groups(my_orthologs)
```

---

compute\_macrosynteny    *Compute significant macrosynteny blocks*

---

### Description

This is a function to generate the contingency table of an orthologs dataframe and apply fisher test to calculate the significant associations. It outputs a dataframe shaped as following : sp1.Chr,sp2.Chr,a,pval,significant,pval\_a

### Usage

```
compute_macrosynteny(orthologs_df, pvalue_threshold = 0.001)
```

### Arguments

orthologs\_df    dataframe. orthologs with genomic coordinates loaded with load\_orthologs()  
pvalue\_threshold    numeric. threshold for significancy. (default equals 0.001)

### Value

A dataframe object

### Examples

```
# basic usage of compute_macrosynteny :  
  
orthologs_table <- system.file("extdata", "my_orthologs.tab", package="macrosyntR")  
  
my_orthologs <- read.table(orthologs_table, header=TRUE)  
  
my_macrosynteny <- compute_macrosynteny(my_orthologs)
```

---

get\_syntenic\_genes    *get the syntenic genes as a table*

---

### Description

This is a function to extract all the syntenic genes from an orthologs\_df. It requires as input an orthologs\_df loaded by load\_orthologs().

### Usage

```
get_syntenic_genes(orthologs_df)
```

**Arguments**

orthologs\_df      dataframe. orthologs with genomic coordinates loaded by load\_orthologs()

**Value**

dataframe composed of details for each detected syntenic block of genes. It contains the following columns : sp1.Chr, sp1.Start, sp1.End, sp2.Chr, sp2.Start, sp2.End, size, sp1.IDs, sp2.IDs

**See Also**

[load\\_orthologs\(\)](#)

**Examples**

```
# basic usage of get_syntenic_genes :
orthologs_table <- system.file("extdata", "my_orthologs.tab", package="macrosyntR")
my_orthologs <- read.table(orthologs_table, header=TRUE)
my_syntenic_block_of_genes <- get_syntenic_genes(my_orthologs)
```

---

load_orthologs	<i>load orthologs with their genomic coordinates.</i>
----------------	---

---

**Description**

Puts together the table of orthologous genes with their genomic coordinates in the two or more species. It outputs a data.frame shaped as following : sp1.ID,sp1.Chr,sp1.Start,sp1.End,sp1.Index,sp2.ID,sp2.Chr,sp2.Start,sp2.Index

**Usage**

```
load_orthologs(
  orthologs_table,
  sp1_bed = NULL,
  sp2_bed = NULL,
  bedfiles = NULL
)
```

**Arguments**

orthologs\_table      character. Full path to the orthologs table (format : geneID\_on\_species1 geneID\_on\_species2 geneID\_on\_speciesN)

sp1\_bed              (deprecated) character. Full path to the genomic coordinates of the genes on species1

sp2_bed	(deprecated) character. Full path to the genomic coordinates of the genes on species2
bedfiles	array. List of full paths to the genomic coordinates ordered as in the appearing order of the orthologs_table (BED format)

**Value**

dataframe composed of genomic coordinates and relative index of orthologs on both species

**Examples**

```
# basic usage of load_orthologs for two species :

orthologs_file <- system.file("extdata", "Bflo_vs_Pyes.tab", package="macrosyntR")
bedfile_sp1 <- system.file("extdata", "Bflo.bed", package="macrosyntR")
bedfile_sp2 <- system.file("extdata", "Pyes.bed", package="macrosyntR")

my_orthologs <- load_orthologs(orthologs_table = orthologs_file,
                              bedfiles = c(bedfile_sp1, bedfile_sp2))

# example with 3 species :
orthologs_file <- system.file("extdata", "Single_copy_orthologs.tsv", package="macrosyntR")
bedfile_sp3 <- system.file("extdata", "Pech.bed", package="macrosyntR")

my_orthologs <- load_orthologs(orthologs_table = orthologs_file,
                              bedfiles = c(bedfile_sp1, bedfile_sp2, bedfile_sp3))
```

---

plot\_chord\_diagram     *plot the Macro-synteny as a chord diagram*

---

**Description**

This is a function to plot the chord diagrams to compare the macro synteny of two or more species. It requires as input an orthologs\_df loaded by load\_orthologs()

**Usage**

```
plot_chord_diagram(
  orthologs_df,
  species_labels = NULL,
  species_labels_size = 5,
  color_by = "sp1.Chr",
  custom_color_palette = NULL,
  reorder_chromosomes = TRUE,
  remove_non_linkage_orthologs = TRUE,
  species_labels_hpos = -400,
```

```

label_size = 2,
ideogram_fill = "white",
ideogram_color = "black",
ideogram_height = 4,
ribbons_curvature = 0.1,
ribbons_alpha = 0.5
)

```

### Arguments

**orthologs\_df** dataframe. orthologs with genomic coordinates loaded by the `load_orthologs()`

**species\_labels** list of characters. names of the species to display on the plot

**species\_labels\_size** integer. size of the labels (default = 2)

**color\_by** string. name of the column in the `orthologs_df` to color the links by (default = "sp1.Chr")

**custom\_color\_palette** list of characters. palette to use for the coloring of the links following the argument `color_by`

**reorder\_chromosomes** logical. (default = TRUE) tells whether to reorder the chromosomes in clusters as implemented in `reorder_macrosynteny()`

**remove\_non\_linkage\_orthologs** logical. (default = TRUE) tells whether to remove the orthologs that are not within significant linkage groups as calculated by `compute_linkage_groups()`.

**species\_labels\_hpos** (default = -400)

**label\_size** integer. size of the labels to display on the ideograms (default = 2)

**ideogram\_fill** character. name of the colors to fill the ideograms with (default = "white")

**ideogram\_color** character. name of the colors to draw the borders of the ideograms with (default = "black")

**ideogram\_height** integer. height of the ideograms (default = 4)

**ribbons\_curvature** float. curvature of the ribbons (default = 0.1)

**ribbons\_alpha** float. alpha of the ribbons (default = 0.5)

### Value

A `ggplot2` object

### See Also

[load\\_orthologs\(\)](#)  
[reorder\\_macrosynteny\(\)](#)  
[compute\\_linkage\\_groups\(\)](#)

**Examples**

```
# basic usage of plot_oxford_grid :  
  
orthologs_table <- system.file("extdata", "my_orthologs.tab", package="macrosyntR")  
  
my_orthologs <- read.table(orthologs_table, header=TRUE)  
  
plot_chord_diagram(my_orthologs, species_labels = c("B. flo", "P. ech"))
```

---

plot\_macrosynteny      *Plot Macro-synteny*

---

**Description**

This is a function to generate the contingency table of an MBH dataframe and apply fisher test to calculate the significant associations.

**Usage**

```
plot_macrosynteny(macrosynt_df, sp1_label = "", sp2_label = "")
```

**Arguments**

macrosynt_df	dataframe of contingency table with p-values calculated by the compute_macrosynteny() function
sp1_label	character. The name of the species1 to display on the plot
sp2_label	character. The name of the species2 to put on the plot

**Value**

ggplot2 object

**See Also**

[compute\\_macrosynteny\(\)](#)

**Examples**

```
# basic usage of plot_macrosynteny :  
  
orthologs_table <- system.file("extdata", "my_orthologs.tab", package="macrosyntR")  
  
my_orthologs <- read.table(orthologs_table, header=TRUE)  
  
my_macrosynteny <- compute_macrosynteny(my_orthologs)  
  
plot_macrosynteny(my_macrosynteny,
```

```
sp1_label = "B. floridae",
sp2_label = "P. yessoensis")
```

---

plot\_oxford\_grid      *plot the Macro-synteny as an oxford grid.*

---

### Description

This is a function to plot the oxford grided plot to compare the macro synteny of two species. It requires as input an orthologs\_df loaded by load\_orthologs()

### Usage

```
plot_oxford_grid(
  orthologs_df,
  sp1_label = "",
  sp2_label = "",
  dot_size = 0.5,
  dot_alpha = 0.4,
  reorder = FALSE,
  keep_only_significant = FALSE,
  color_by = NULL,
  pvalue_threshold = 0.001,
  color_palette = NULL,
  shade_non_significant = TRUE,
  reverse_species = FALSE,
  keep_sp1_raw_order = FALSE
)
```

### Arguments

orthologs_df	dataframe. orthologs with genomic coordinates loaded by the load_orthologs()
sp1_label	character. name of 1st species to display on the plot
sp2_label	character. name of 2nd species to display on the plot
dot_size	numeric. (default = 0.5)
dot_alpha	numeric. (default = 0.4)
reorder	logical. (default = FALSE) tells whether to reorder the chromosomes in clusters as implemented in reorder_macrosynteny()
keep_only_significant	logical. (default = FALSE)
color_by	string/variable name. (default = NULL) column of the orthologs_df to use to color the dots.
pvalue_threshold	numeric. (default = 0.001)



`color_palette` vector. (default = NULL) list of colors (as string under double quote) for the clusters. The amount of colors must match the amount of clusters.

`shade_non_significant` logical. (default = TRUE) When TRUE the orthologs located on non-significant linkage groups are displayed in "grey"

`reverse_species` logical. (default = FALSE) When TRUE the x and y axis of the plot are reversed. sp1 is displayed on the y axis and sp2 is displayed on the x axis.

`keep_sp1_raw_order` logical.(default equals FALSE) tells if the reordering should be constrained on the species1 and change just the order of the species2

**Value**

A ggplot2 object

**See Also**

[load\\_orthologs\(\)](#)

[reorder\\_macrosynteny\(\)](#)

**Examples**

```
# basic usage of plot_oxford_grid :

orthologs_table <- system.file("extdata", "my_orthologs.tab", package="macrosyntR")

my_orthologs <- read.table(orthologs_table, header=TRUE)

plot_oxford_grid(my_orthologs,
                 sp1_label = "B. floridae",
                 sp2_label = "P. echinospica")

# plot a reordered Oxford Grid and color by cluster :

plot_oxford_grid(my_orthologs,
                 sp1_label = "B. floridae",
                 sp2_label = "P. echinospica",
                 reorder = TRUE,
                 color_by = "clust")
```

---

reorder\_macrosynteny *Reorder the mbh\_df before plotting*

---

**Description**

This is a function to reorder an orthologs\_df, that was generated with load\_orthologs(). It retrieves communities using igraph::cluster\_fast\_greedy.

**Usage**

```
reorder_macrosynteny(  
  orthologs_df,  
  pvalue_threshold = 0.001,  
  keep_only_significant = FALSE,  
  keep_sp1_raw_order = FALSE  
)
```

**Arguments**

`orthologs_df`    dataframe. mutual best hits with genomic coordinates loaded with `load_orthologs()`

`pvalue_threshold`  
                  numeric. threshold for significancy. (default equals 0.001)

`keep_only_significant`  
                  logical. (default equals FALSE) tells if the non significant linkage groups should be removed. It drastically speeds up the computation when using one highly fragmented genome.

`keep_sp1_raw_order`  
                  logical. (default equals FALSE) tells if the reordering should be constrained on the species1 and change just the order of the species2

**Value**

A dataframe object

**See Also**

[load\\_orthologs\(\)](#)  
[compute\\_macrosynteny\(\)](#)

**Examples**

```
# basic usage of reorder_macrosynteny :  
  
orthologs_table <- system.file("extdata", "my_orthologs.tab", package="macrosyntR")  
  
my_orthologs <- read.table(orthologs_table, header=TRUE)  
  
my_orthologs_reordered <- reorder_macrosynteny(my_orthologs)
```

---

`reorder_multiple_macrosyntenies`*Reorder the chromosomes of two or more species before plotting*

---

## Description

This is a function to reorder an `orthologs_df`, same as `reorder_macrosyteny`, but it handles tables with more than 2 species.

## Usage

```
reorder_multiple_macrosyntenies(orthologs_df)
```

## Arguments

`orthologs_df`    dataframe. orthologs with genomic coordinates loaded with `load_orthologs()`

## Value

A dataframe object

## See Also

[load\\_orthologs\(\)](#)

[compute\\_macrosyteny\(\)](#)

[reorder\\_macrosyteny\(\)](#)

## Examples

```
# basic usage of reorder_macrosyteny :  
orthologs_table <- system.file("extdata", "my_orthologs.tab", package="macrosyntR")  
my_orthologs <- read.table(orthologs_table, header=TRUE)  
my_orthologs_reordered <- reorder_multiple_macrosyntenies(my_orthologs)
```

---

reverse\_species\_order *Reverse order of the species in an orthologs\_df.*

---

### Description

Returns an orthologs\_df (data.frame) with reversed species order compared to the inputted orthologs\_df. sp1 becomes sp2 and the otherway around. It intends at facilitating the integration of more than just two datasets. It outputs a data.frame shaped as following : sp1.ID,sp1.Chr,sp1.Start,sp1.End,sp1.Index,sp2.ID,

### Usage

```
reverse_species_order(orthologs_df)
```

### Arguments

orthologs\_df    orthologs\_df dataframe. mutual best hits with genomic coordinates loaded with load\_orthologs()

### Value

dataframe composed of genomic coordinates and relative index of orthologs on both species

### See Also

[load\\_orthologs\(\)](#)

### Examples

```
# basic usage of reverse_species_order :

orthologs_table <- system.file("extdata", "my_orthologs.tab", package="macrosyntR")

my_orthologs <- read.table(orthologs_table, header=TRUE)

my_orthologs_reversed <- reverse_species_order(my_orthologs)
```

---

subset\_linkage\_orthologs

*Subset Orthologs contained in conserved linkage groups*

---

### Description

This is a function to subset an orthologs\_df and keep only the orthologs that are within significant linkage groups computed by the function compute\_linkage\_groups().

**Usage**

```
subset_linkage_orthologs(orthologs_df, linkages = NULL)
```

**Arguments**

<code>orthologs_df</code>	dataframe. orthologs with genomic coordinates loaded with <code>load_orthologs()</code>
<code>linkages</code>	dataframe. table listing the linkage groups as returned by the function <code>compute_linkage_groups()</code>

**Value**

A dataframe object

**See Also**

[load\\_orthologs\(\)](#)

[compute\\_linkage\\_groups\(\)](#)

**Examples**

```
# basic usage of compute_linkage_groups:  
  
orthologs_table <- system.file("extdata", "my_orthologs.tab", package="macrosyntR")  
  
my_orthologs <- read.table(orthologs_table, header=TRUE)  
  
my_macrosynteny <- compute_linkage_groups(my_orthologs)
```

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