

Package ‘zinbwave’

October 22, 2025

Type Package

Title Zero-Inflated Negative Binomial Model for RNA-Seq Data

Version 1.31.0

Description Implements a general and flexible zero-inflated negative binomial model that can be used to provide a low-dimensional representations of single-cell RNA-seq data. The model accounts for zero inflation (dropouts), over-dispersion, and the count nature of the data. The model also accounts for the difference in library sizes and optionally for batch effects and/or other covariates, avoiding the need for pre-normalize the data.

License Artistic-2.0

Depends R (>= 3.4), methods, SummarizedExperiment,
SingleCellExperiment

Imports BiocParallel, softImpute, stats, genefilter, edgeR, Matrix

Suggests knitr, rmarkdown, testthat, matrixStats, magrittr, scRNAseq,
ggplot2, biomaRt, BiocStyle, Rtsne, DESeq2, sparseMatrixStats

VignetteBuilder knitr

LazyData TRUE

RoxygenNote 7.3.1

biocViews ImmunoOncology, DimensionReduction, GeneExpression, RNASeq,
Software, Transcriptomics, Sequencing, SingleCell

BugReports <https://github.com/drisso/zinbwave/issues>

git_url <https://git.bioconductor.org/packages/zinbwave>

git_branch devel

git_last_commit c6e6f01

git_last_commit_date 2025-04-15

Repository Bioconductor 3.22

Date/Publication 2025-10-21

Author Davide Risso [aut, cre, cph],
Svetlana Gribkova [aut],
Fanny Perraudeau [aut],
Jean-Philippe Vert [aut],
Clara Bagatin [aut]

Maintainer Davide Risso <risso.davide@gmail.com>

Contents

computeDevianceResiduals	3
computeObservationalWeights	4
getAlpha_mu	4
getAlpha_pi	5
getBeta_mu	5
getBeta_pi	6
getEpsilon_alpha	6
getEpsilon_beta_mu	7
getEpsilon_beta_pi	7
getEpsilon_gamma_mu	8
getEpsilon_gamma_pi	8
getEpsilon_W	9
getEpsilon_zeta	9
getGamma_mu	10
getGamma_pi	10
getLogitPi	11
getLogMu	12
getMu	12
getPhi	13
getPi	13
getTheta	14
getV_mu	15
getV_pi	15
getW	16
getX_mu	16
getX_pi	17
getZeta	17
glmWeightedF	18
imputeZeros	19
independentFiltering	20
loglik	20
nFactors	21
nFeatures	22
nParams	22
nSamples	23
orthogonalizeTraceNorm	23
penalty	24
pvalueAdjustment	25
solveRidgeRegression	26
toydata	27
zinb.loglik	27
zinb.loglik.dispersion	28
zinb.loglik.dispersion.gradient	29
zinb.loglik.matrix	29
zinb.loglik.regression	30
zinb.loglik.regression.gradient	31
zinb.regression.parseModel	32
zinbAIC	33
zinbFit	34
zinbInitialize	36

<i>computeDevianceResiduals</i>	3
zinbModel	37
ZinbModel-class	39
zinbOptimize	43
zinbOptimizeDispersion	44
zinbSim	45
zinbsurf	46
zinbwave	47
Index	50

`computeDevianceResiduals`

Deviance residuals of the zero-inflated negative binomial model

Description

Given a matrix of counts, this function computes the deviance residuals under a zero-inflated negative binomial (ZINB) model.

Usage

```
computeDevianceResiduals(model, x, ignoreW = TRUE)
```

Arguments

<code>model</code>	the zinb model
<code>x</code>	the matrix of counts n cells by J genes
<code>ignoreW</code>	logical, if true matrix W is ignored. Default is TRUE.

Value

the matrix of deviance residuals of the model.

Examples

```
se <- SummarizedExperiment(matrix(rpois(60, lambda=5), nrow=10, ncol=6),
  colData = data.frame(bio = gl(2, 3)))
m <- zinbFit(se, X=model.matrix(~bio, data=colData(se)),
  BPPARAM=BiocParallel::SerialParam())
computeDevianceResiduals(m, t(assay(se)))
```

```
computeObservationalWeights
```

Observational weights of the zero-inflated negative binomial model for each entry in the matrix of counts

Description

Given a matrix of counts, this function computes the observational weights of the counts under a zero-inflated negative binomial (ZINB) model. For each count, the ZINB distribution is parametrized by three parameters: the mean value and the dispersion of the negative binomial distribution, and the probability of the zero component.

Usage

```
computeObservationalWeights(model, x)
```

Arguments

model	the zinb model
x	the matrix of counts

Value

the matrix of observational weights computed from the model.

Examples

```
se <- SummarizedExperiment(matrix(rpois(60, lambda=1), nrow=10, ncol=6),
  colData = data.frame(bio = gl(2, 3)))
m <- zinbFit(se, X=model.matrix(~bio, data=colData(se)),
  BPPARAM=BiocParallel::SerialParam())
computeObservationalWeights(m, assay(se))
```

```
getAlpha_mu
```

Returns the matrix of parameters alpha_mu

Description

Given an object that describes a matrix of zero-inflated distributions, returns the matrix of parameters associated with W for the mean part (mu)

Usage

```
getAlpha_mu(object, ...)
```

Arguments

object	an object that describes a matrix of zero-inflated distributions.
...	Additional parameters.

Value

the matrix of alpha_mu parameters

Examples

```
a <- zinbModel(n=5, J=10)
getAlpha_mu(a)
```

getAlpha_pi	<i>Returns the matrix of paramters alpha_pi</i>
-------------	---

Description

Given an object that describes a matrix of zero-inflated distributions, returns the matrix of parameters associated with W for the zero part (pi)

Usage

```
getAlpha_pi(object, ...)
```

Arguments

object	an object that describes a matrix of zero-inflated distributions.
...	Additional parameters.

Value

the matrix of alpha_pi parameters

Examples

```
a <- zinbModel(n=5, J=10)
getAlpha_pi(a)
```

getBeta_mu	<i>Returns the matrix of paramters beta_mu</i>
------------	--

Description

Given an object that describes a matrix of zero-inflated distributions, returns the matrix of parameters associated with X_mu

Usage

```
getBeta_mu(object, ...)
```

Arguments

object	an object that describes a matrix of zero-inflated distributions.
...	Additional parameters.

Value

the matrix of beta_mu parameters

Examples

```
a <- zinbModel(n=5, J=10)
getBeta_mu(a)
```

getBeta_pi

Returns the matrix of parameters beta_pi

Description

Given an object that describes a matrix of zero-inflated distributions, returns the matrix of parameters associated with X_pi

Usage

```
getBeta_pi(object, ...)
```

Arguments

object an object that describes a matrix of zero-inflated distributions.
 ... Additional parameters.

Value

the matrix of beta_pi parameters

Examples

```
a <- zinbModel(n=5, J=10)
getBeta_pi(a)
```

getEpsilon_alpha

Returns the vector of regularization parameter for alpha

Description

Given an object describing a ZINB model, returns a vector of size the number of rows in the parameter alpha with the regularization parameters associated to each row. Here alpha refers to both alpha_mu and alpha_pi, which have the same size and have the same regularization.

Usage

```
getEpsilon_alpha(object)
```

Arguments

object an object that describes a matrix of zero-inflated distributions.

Value

the regularization parameters for alpha_mu and alpha_pi.

Examples

```
a <- zinbModel(n=5, J=10)
getEpsilon_alpha(a)
```

getEpsilon_beta_mu	Returns the vector of regularization parameter for beta_mu
--------------------	--

Description

Given an object describing a ZINB model, returns a vector of size the number of rows in the parameter beta_mu with the regularization parameters associated to each row.

Usage

```
getEpsilon_beta_mu(object)
```

Arguments

object an object that describes a matrix of zero-inflated distributions.

Value

the regularization parameters for beta_mu.

Examples

```
a <- zinbModel(n=5, J=10)
getEpsilon_beta_mu(a)
```

getEpsilon_beta_pi	Returns the vector of regularization parameter for beta_pi
--------------------	--

Description

Given an object describing a ZINB model, returns a vector of size the number of rows in the parameter beta_pi with the regularization parameters associated to each row.

Usage

```
getEpsilon_beta_pi(object)
```

Arguments

object an object that describes a matrix of zero-inflated distributions.

Value

the regularization parameters for beta_pi.

Examples

```
a <- zinbModel(n=5, J=10)
getEpsilon_beta_pi(a)
```

getEpsilon_gamma_mu	<i>Returns the vector of regularization parameter for gamma_mu</i>
---------------------	--

Description

Given an object describing a ZINB model, returns a vector of size the number of columns in the parameter gamma_mu with the regularization parameters associated to each row.

Usage

```
getEpsilon_gamma_mu(object)
```

Arguments

object an object that describes a matrix of zero-inflated distributions.

Value

the regularization parameters for gamma_mu.

Examples

```
a <- zinbModel(n=5, J=10)
getEpsilon_gamma_mu(a)
```

getEpsilon_gamma_pi	<i>Returns the vector of regularization parameter for gamma_pi</i>
---------------------	--

Description

Given an object describing a ZINB model, returns a vector of size the number of columns in the parameter gamma_pi with the regularization parameters associated to each column.

Usage

```
getEpsilon_gamma_pi(object)
```

Arguments

object an object that describes a matrix of zero-inflated distributions.

Value

the regularization parameters for gamma_pi.

Examples

```
a <- zinbModel(n=5, J=10)
getEpsilon_gamma_pi(a)
```

getEpsilon_W

Returns the vector of regularization parameter for W

Description

Given an object describing a ZINB model, returns a vector of size the number of columns in the parameter W with the regularization parameters associated to each column.

Usage

```
getEpsilon_W(object)
```

Arguments

object an object that describes a matrix of zero-inflated distributions.

Value

the regularization parameters for W.

Examples

```
a <- zinbModel(n=5, J=10)
getEpsilon_W(a)
```

getEpsilon_zeta

Returns the regularization parameter for the dispersion parameter

Description

The regularization parameter penalizes the variance of zeta, the log of the dispersion parameters across samples.

Usage

```
getEpsilon_zeta(object)
```

Arguments

object an object that describes a matrix of zero-inflated distributions.

Value

the regularization parameters for zeta.

Examples

```
a <- zinbModel(n=5, J=10)
getEpsilon_zeta(a)
```

getGamma_mu	<i>Returns the matrix of paramters gamma_mu</i>
-------------	---

Description

Given an object that describes a matrix of zero-inflated distributions, returns the matrix of parameters associated with V_{μ}

Usage

```
getGamma_mu(object, ...)
```

Arguments

object	an object that describes a matrix of zero-inflated distributions.
...	Additional parameters.

Value

the matrix of gamma_mu parameters

Examples

```
a <- zinbModel(n=5, J=10)
getGamma_mu(a)
```

getGamma_pi	<i>Returns the matrix of paramters gamma_pi</i>
-------------	---

Description

Given an object that describes a matrix of zero-inflated distributions, returns the matrix of parameters associated with V_{π}

Usage

```
getGamma_pi(object, ...)
```

Arguments

object	an object that describes a matrix of zero-inflated distributions.
...	Additional parameters.

Value

the matrix of gamma_pi parameters

Examples

```
a <- zinbModel(n=5, J=10)
getGamma_pi(a)
```

getLogitPi

Returns the matrix of logit of probabilities of zero

Description

Given an object that describes a matrix of zero-inflated distributions, returns the matrix of logit of probabilities of 0.

Usage

```
getLogitPi(object)
```

Arguments

`object` an object that describes a matrix of zero-inflated distributions.

Details

Note that although the user interface of [zinbFit](#) requires a J x n matrix, internally this is stored as a n x J matrix (i.e., samples in row and genes in column). Hence the parameter matrix returned by this function is of n x J dimensions.

Value

the matrix of logit-probabilities of 0

Examples

```
a <- zinbModel(n=5, J=10)
getLogitPi(a)
```

getLogMu

Returns the matrix of logarithm of mean parameters

Description

Given an object that describes a matrix of zero-inflated distributions, returns the matrix of logarithm of mean parameters.

Usage

```
getLogMu(object)
```

Arguments

object an object that describes a matrix of zero-inflated distributions.

Details

Note that although the user interface of [zinbFit](#) requires a $J \times n$ matrix, internally this is stored as a $n \times J$ matrix (i.e., samples in row and genes in column). Hence the parameter matrix returned by this function is of $n \times J$ dimensions.

Value

the matrix of logarithms of mean parameters

Examples

```
a <- zinbModel(n=5, J=10)
getLogMu(a)
```

getMu

Returns the matrix of mean parameters

Description

Given an object that describes a matrix of zero-inflated distributions, returns the matrix of mean parameters.

Usage

```
getMu(object)
```

Arguments

object an object that describes a matrix of zero-inflated distributions.

Details

Note that although the user interface of [zinbFit](#) requires a $J \times n$ matrix, internally this is stored as a $n \times J$ matrix (i.e., samples in row and genes in column). Hence the parameter matrix returned by this function is of $n \times J$ dimensions.

Value

the matrix of mean parameters

Examples

```
a <- zinbModel(n=5, J=10)
getMu(a)
```

getPhi

Returns the vector of dispersion parameters

Description

Given an object that describes a matrix of zero-inflated negative binomial distributions, returns the vector of dispersion parameters `phi`.

Usage

```
getPhi(object)
```

Arguments

`object` an object that describes a matrix of zero-inflated. distributions.

Value

the vector of dispersion parameters

Examples

```
a <- zinbModel(n=5, J=10)
getPhi(a)
```

getPi

Returns the matrix of probabilities of zero

Description

Given an object that describes a matrix of zero-inflated distributions, returns the matrix of probabilities of 0.

Usage

```
getPi(object)
```

Arguments

`object` an object that describes a matrix of zero-inflated distributions.

Details

Note that although the user interface of `zinbFit` requires a $J \times n$ matrix, internally this is stored as a $n \times J$ matrix (i.e., samples in row and genes in column). Hence the parameter matrix returned by this function is of $n \times J$ dimensions.

Value

the matrix of probabilities of 0

Examples

```
a <- zinbModel(n=5, J=10)
getPi(a)
```

`getTheta`*Returns the vector of inverse dispersion parameters*

Description

Given an object that describes a matrix of zero-inflated negative binomial distributions, returns the vector of inverse dispersion parameters `theta`.

Usage

```
getTheta(object)
```

Arguments

`object` an object that describes a matrix of zero-inflated distributions.

Value

the vector of inverse dispersion parameters `theta`

Examples

```
a <- zinbModel(n=5, J=10)
getTheta(a)
```

getV_mu	Returns the gene-level design matrix for mu
---------	---

Description

Given an object that describes a matrix of zero-inflated distributions, returns the gene-level design matrix for mu

Usage

```
getV_mu(object, ...)
```

Arguments

object	an object that describes a matrix of zero-inflated distributions.
...	Additional parameters.

Value

the gene-level design matrix for mu

Examples

```
a <- zinbModel(n=5, J=10)
getV_mu(a)
```

getV_pi	Returns the gene-level design matrix for pi
---------	---

Description

Given an object that describes a matrix of zero-inflated distributions, returns the gene-level design matrix for pi

Usage

```
getV_pi(object, ...)
```

Arguments

object	an object that describes a matrix of zero-inflated distributions.
...	Additional parameters.

Value

the gene-level design matrix for pi

Examples

```
a <- zinbModel(n=5, J=10)
getV_pi(a)
```

getW	<i>Returns the low-dimensional matrix of inferred sample-level covariates W</i>
------	--

Description

Given an object that contains the fit of a ZINB-WaVE model, returns the matrix W of low-dimensional matrix of inferred sample-level covariates.

Usage

```
getW(object)
```

Arguments

object a [ZinbModel](#) object, typically the result of [zinbFit](#).

Value

the matrix W of inferred sample-level covariates.

Examples

```
a <- zinbModel(n=5, J=10)
getW(a)
```

getX_mu	<i>Returns the sample-level design matrix for μ</i>
---------	--

Description

Given an object that describes a matrix of zero-inflated distributions, returns the sample-level design matrix for μ

Usage

```
getX_mu(object, ...)
```

Arguments

object an object that describes a matrix of zero-inflated distributions.
... Additional parameters.

Value

the sample-level design matrix for μ

Examples

```
a <- zinbModel(n=5, J=10)
getX_mu(a)
```

getX_pi	Returns the sample-level design matrix for pi
---------	---

Description

Given an object that describes a matrix of zero-inflated distributions, returns the sample-level design matrix for pi

Usage

```
getX_pi(object, ...)
```

Arguments

object	an object that describes a matrix of zero-inflated distributions.
...	Additional parameters.

Value

the sample-level design matrix for pi

Examples

```
a <- zinbModel(n=5, J=10)
getX_pi(a)
```

getZeta	Returns the vector of log of inverse dispersion parameters
---------	--

Description

Given an object that describes a matrix of zero-inflated negative binomial distributions, returns the vector zeta of log of inverse dispersion parameters

Usage

```
getZeta(object)
```

Arguments

object	an object that describes a matrix of zero-inflated distributions.
--------	---

Value

the vector zeta of log of inverse dispersion parameters

Examples

```
a <- zinbModel(n=5, J=10)
getZeta(a)
```

glmWeightedF	<i>Zero-inflation adjusted statistical tests for assessing differential expression.</i>
--------------	---

Description

This function recycles an old version of the [glmLRT](#) method that allows an F-test with adjusted denominator degrees of freedom to account for the downweighting in the zero-inflation model.

Usage

```
glmWeightedF(
  glmfit,
  coef = ncol(glmfit$design),
  contrast = NULL,
  ZI = TRUE,
  independentFiltering = TRUE,
  filter = NULL
)
```

Arguments

glmfit	a DGEGLM-class object, usually output from glmFit .
coef	integer or character vector indicating which coefficients of the linear model are to be tested equal to zero. Values must be columns or column names of design. Defaults to the last coefficient. Ignored if contrast is specified.
contrast	numeric vector or matrix specifying one or more contrasts of the linear model coefficients to be tested equal to zero. Number of rows must equal to the number of columns of design. If specified, then takes precedence over coef.
ZI	logical, specifying whether the degrees of freedom in the statistical test should be adjusted according to the weights in the fit object to account for the downweighting. Defaults to TRUE and this option is highly recommended.
independentFiltering	logical, specifying whether independent filtering should be performed.
filter	vector of values to perform filtering on. Default is the mean of the fitted values from glmfit.

Details

When ‘independentFiltering=TRUE’ (default) an independent filtering step is applied prior to the multiple testing procedure, as described in great details in the ‘DESeq2’ vignette. The values in the ‘padjFilter’ column refer to this procedure. They are identical to the ‘FDR’ values if the filtering step does not remove any gene, since the function uses the Benjamini-Hochberg correction by default. If the procedure filters some genes, the adjusted p-values will typically result in greater power to detect DE genes. The theory behind independent filtering is described in Bourgon et al. (2010).

Note

This function uses an adapted version of the [glmLRT](#) function that was originally written by Gordon Smyth, Davis McCarthy and Yunshun Chen as part of the edgeR package. Koen Van den Berge wrote code to adjust residual degree of freedom and added the independent filtering step.

References

McCarthy, DJ, Chen, Y, Smyth, GK (2012). Differential expression analysis of multifactor RNA-Seq experiments with respect to biological variation. *Nucleic Acids Research* 40, 4288-4297. Bourgon, Richard, Robert Gentleman, and Wolfgang Huber (2010) Independent Filtering Increases Detection Power for High-Throughput Experiments. *PNAS* 107 (21): 9546-51.

See Also

[glmLRT](#)

imputeZeros	<i>Impute the zeros using the estimated parameters from the ZINB model.</i>
-------------	---

Description

Given a matrix of counts and a zinb model, this function computes the imputed counts under a zero-inflated negative binomial (ZINB) model.

Usage

```
imputeZeros(model, x)
```

Arguments

model	the zinb model
x	the matrix of counts n cells by J genes

Value

the matrix of imputed counts.

Examples

```
se <- SummarizedExperiment(matrix(rpois(60, lambda=5), nrow=10, ncol=6),
  colData = data.frame(bio = gl(2, 3)))
m <- zinbFit(se, X=model.matrix(~bio, data=colData(se)),
  BPPARAM=BiocParallel::SerialParam())
imputeZeros(m, t(assay(se)))
```

independentFiltering	<i>Perform independent filtering in differential expression analysis.</i>
----------------------	---

Description

This function uses the DESeq2 independent filtering method to increase detection power in high throughput gene expression studies.

Usage

```
independentFiltering(object, filter, objectType = c("edgeR", "limma"))
```

Arguments

object	Either a DGELRT-class object or a data.frame with differential expression results.
filter	The characteristic to use for filtering, usually a measure of normalized mean expression for the features.
objectType	Either "edgeR" or "limma". If "edgeR", it is assumed that object is of class DGELRT-class , the output of glmLRT . If "limma", it is assumed that object is a data.frame and the output of a limma-voom analysis.

Author(s)

Koen Van den Berge

References

Michael I Love, Wolfgang Huber, and Simon Anders. Moderated estimation of fold change and dispersion for RNA-seq data with DESeq2. *Genome Biology*, 15(12):550, dec 2014.

See Also

[results](#)

loglik	<i>Compute the log-likelihood of a model given some data</i>
--------	--

Description

Given a statistical model and some data, this function computes the log-likelihood of the model given the data, i.e., the log-probability of the data under the model.

Usage

```
loglik(model, x, ...)

## S4 method for signature 'ZinbModel,matrix'
loglik(model, x)
```

Arguments

<code>model</code>	an object that describes a statistical model.
<code>x</code>	an object that describes data.
<code>...</code>	additional arguments.

Value

The log-likelihood of the model given the data.

Methods (by class)

- `loglik(model = ZinbModel, x = matrix)`: return the log-likelihood of the ZINB model.

Examples

```
m <- zinbModel(n=5, J=10)
x <- zinbSim(m)
loglik(m, x$counts)
```

nFactors

Generic function that returns the number of latent factors

Description

Given an object that describes a dataset or a model involving latent factors, this function returns the number of latent factors.

Usage

```
nFactors(x)
```

Arguments

<code>x</code>	an object that describes a dataset or a model involving latent factors
----------------	--

Value

the number of latent factors

Examples

```
a <- zinbModel()
nFactors(a)
```

nFeatures

Generic function that returns the number of features

Description

Given an object that describes a dataset or a model, it returns the number of features.

Usage

```
nFeatures(x)
```

Arguments

x an object that describes a dataset or a model.

Value

the number of features.

Examples

```
a <- zinbModel()
nFeatures(a)
```

nParams

Generic function that returns the total number of parameters of the model

Description

Given an object that describes a model or a dataset, it returns total number of parameters of the model.

Usage

```
nParams(model)
```

```
## S4 method for signature 'ZinbModel'
nParams(model)
```

Arguments

model an object that describes a dataset or a model.

Value

the total number of parameters of the model.

Functions

- nParams(ZinbModel): returns the total number of parameters in the model.

Examples

```
a <- zinbModel()
nParams(a)
```

nSamples

*Generic function that returns the number of samples***Description**

Given an object that describes a model or a dataset, it returns the number of samples.

Usage

```
nSamples(x)
```

Arguments

x an object that describes a dataset or a model.

Value

the number of samples.

Examples

```
a <- zinbModel()
nSamples(a)
```

orthogonalizeTraceNorm

*Orthogonalize matrices to minimize trace norm of their product***Description**

Given two matrices U and V that can be multiplied, this function finds two new matrices U2 and V2 such that their product is conserved ($U \cdot V = U2 \cdot V2$) and such that $a\|U\|^2 + b\|V\|^2$ is minimized.

Usage

```
orthogonalizeTraceNorm(U, V, a = 1, b = 1)
```

Arguments

U	left matrix
V	right matrix
a	weight of the norm of U (default=1)
b	weight of the norm of V (default=1)

Value

A list with the two matrices that solve the problem in the slots U and V.

Examples

```
U <- matrix(rnorm(15),5,3)
V <- matrix(rnorm(12),3,4)
o <- orthogonalizeTraceNorm(U,V)
norm( U%*%V - o$U%*%o$V) # should be zero
sum(U^2)+sum(V^2)
sum(o$U^2)+sum(o$V^2) # should be smaller
```

penalty	<i>Compute the penalty of a model</i>
---------	---------------------------------------

Description

Given a statistical model with regularization parameters, compute the penalty.

Usage

```
penalty(model)

## S4 method for signature 'ZinbModel'
penalty(model)
```

Arguments

`model` an object that describes a statistical model with regularization parameters.

Value

The penalty of the model.

Methods (by class)

- `penalty(ZinbModel)`: return the penalization.

Examples

```
m <- zinbModel(K=2)
penalty(m)
```

pvalueAdjustment	<i>Perform independent filtering in differential expression analysis.</i>
------------------	---

Description

This function performs independent filtering to increase detection power in high throughput gene expression studies.

Usage

```
pvalueAdjustment(  
  baseMean,  
  filter,  
  pValue,  
  theta,  
  alpha = 0.05,  
  pAdjustMethod = "BH"  
)
```

Arguments

baseMean	A vector of mean values.
filter	A vector of stage-one filter statistics.
pValue	A vector of unadjusted p-values, or a function which is able to compute this vector from the filtered portion of data, if data is supplied. The option to supply a function is useful when the value of the test statistic depends on which hypotheses are filtered out at stage one. (The limma t-statistic is an example.)
theta	A vector with one or more filtering fractions to consider. Actual cutoffs are then computed internally by applying quantile to the filter statistics contained in (or produced by) the filter argument.
alpha	A cutoff to which p-values, possibly adjusted for multiple testing, will be compared. Default is 0.05.
pAdjustMethod	The unadjusted p-values contained in (or produced by) test will be adjusted for multiple testing after filtering. Default is "BH".

Value

a list with pvalues, filtering threshold, theta, number of rejections, and alpha.

Note

This function is an adapted version of the pvalueAdjustment function that was originally written by Michael I. Love as part of the DESeq2 package. Koen Van den Berge adapted the function.

`solveRidgeRegression` *Solve ridge regression or logistic regression problems*

Description

This function solves a regression or logistic regression problem regularized by a L2 or weighted L2 penalty. Contrary to `lm.ridge` or `glmnet`, it works for any number of predictors.

Usage

```
solveRidgeRegression(
  x,
  y,
  beta = rep(0, NCOL(x)),
  epsilon = 1e-06,
  family = c("gaussian", "binomial"),
  offset = rep(0, NROW(x))
)
```

Arguments

<code>x</code>	a matrix of covariates, one sample per row, one covariate per column.
<code>y</code>	a vector of response (continuous for regression, 0/1 binary for logistic regression)
<code>beta</code>	an initial solution where optimization starts (null vector by default)
<code>epsilon</code>	a scalar or vector of regularization parameters (default 1e-6)
<code>family</code>	a string to choose the type of regression (default family="gaussian")
<code>offset</code>	a vector of offsets (default null vector)

Details

When family="gaussian", we solve the ridge regression problem that finds the β that minimizes:

$$\|y - x\beta\|^2 + \epsilon\|\beta\|^2/2.$$

When family="binomial" we solve the ridge logistic regression problem

$$\min \sum_i [-y_i(x\beta)_i + \log(1 + \exp(x\beta)_i))] + \epsilon\|\beta\|^2/2.$$

When epsilon is a vector of size equal to the size of beta, then the penalty is a weighted L2 norm $\sum_i \epsilon_i \beta_i^2/2$.

Value

A vector solution of the regression problem

toydata	<i>Toy dataset to check the model</i>
---------	---------------------------------------

Description

Toy dataset to check the model

Format

A matrix of integers (counts) with 96 samples (rows) and 500 genes (columns).

zinb.loglik	<i>Log-likelihood of the zero-inflated negative binomial model</i>
-------------	--

Description

Given a vector of counts, this function computes the sum of the log-probabilities of the counts under a zero-inflated negative binomial (ZINB) model. For each count, the ZINB distribution is parametrized by three parameters: the mean value and the dispersion of the negative binomial distribution, and the probability of the zero component.

Usage

```
zinb.loglik(Y, mu, theta, logitPi)
```

Arguments

Y	the vector of counts
mu	the vector of mean parameters of the negative binomial
theta	the vector of dispersion parameters of the negative binomial, or a single scalar is also possible if the dispersion parameter is constant. Note that theta is sometimes called inverse dispersion parameter (and $\phi=1/\theta$ is then called the dispersion parameter). We follow the convention that the variance of the NB variable with mean μ and dispersion θ is $\mu + \mu^2/\theta$.
logitPi	the vector of logit of the probabilities of the zero component

Value

the log-likelihood of the model.

Examples

```
n <- 10
mu <- seq(10,50,length.out=n)
logitPi <- rnorm(10)
zeta <- rnorm(10)
Y <- rnbino(n=n, size=exp(zeta), mu=mu)
zinb.loglik(Y, mu, exp(zeta), logitPi)
zinb.loglik(Y, mu, 1, logitPi)
```

`zinb.loglik.dispersion`*Log-likelihood of the zero-inflated negative binomial model, for a fixed dispersion parameter*

Description

Given a unique dispersion parameter and a set of counts, together with a corresponding set of mean parameters and probabilities of zero components, this function computes the sum of the log-probabilities of the counts under the ZINB model. The dispersion parameter is provided to the function through `zeta = log(theta)`, where `theta` is sometimes called the inverse dispersion parameter. The probabilities of the zero components are provided through their logit, in order to better numerical stability.

Usage

```
zinb.loglik.dispersion(zeta, Y, mu, logitPi)
```

Arguments

<code>zeta</code>	a scalar, the log of the inverse dispersion parameters of the negative binomial model
<code>Y</code>	a vector of counts
<code>mu</code>	a vector of mean parameters of the negative binomial
<code>logitPi</code>	a vector of logit of the probabilities of the zero component

Value

the log-likelihood of the model.

See Also

[zinb.loglik.](#)

Examples

```
mu <- seq(10,50,length.out=10)
logitPi <- rnorm(10)
zeta <- rnorm(10)
Y <- rnbinom(n=10, size=exp(zeta), mu=mu)
zinb.loglik.dispersion(zeta, Y, mu, logitPi)
```

zinb.loglik.dispersion.gradient

Derivative of the log-likelihood of the zero-inflated negative binomial model with respect to the log of the inverse dispersion parameter

Description

Derivative of the log-likelihood of the zero-inflated negative binomial model with respect to the log of the inverse dispersion parameter

Usage

```
zinb.loglik.dispersion.gradient(zeta, Y, mu, logitPi)
```

Arguments

zeta	the log of the inverse dispersion parameters of the negative binomial
Y	a vector of counts
mu	a vector of mean parameters of the negative binomial
logitPi	a vector of the logit of the probability of the zero component

Value

the gradient of the inverse dispersion parameters.

See Also

[zinb.loglik](#), [zinb.loglik.dispersion](#).

zinb.loglik.matrix

Log-likelihood of the zero-inflated negative binomial model for each entry in the matrix of counts

Description

Given a matrix of counts, this function computes the log-probabilities of the counts under a zero-inflated negative binomial (ZINB) model. For each count, the ZINB distribution is parametrized by three parameters: the mean value and the dispersion of the negative binomial distribution, and the probability of the zero component.

Usage

```
zinb.loglik.matrix(model, x)
```

Arguments

model	the zinb model
x	the matrix of counts

Value

the matrix of log-likelihood of the model.

```
zinb.loglik.regression
```

Penalized log-likelihood of the ZINB regression model

Description

This function computes the penalized log-likelihood of a ZINB regression model given a vector of counts.

Usage

```
zinb.loglik.regression(
  alpha,
  Y,
  A.mu = matrix(nrow = length(Y), ncol = 0),
  B.mu = matrix(nrow = length(Y), ncol = 0),
  C.mu = matrix(0, nrow = length(Y), ncol = 1),
  A.pi = matrix(nrow = length(Y), ncol = 0),
  B.pi = matrix(nrow = length(Y), ncol = 0),
  C.pi = matrix(0, nrow = length(Y), ncol = 1),
  C.theta = matrix(0, nrow = length(Y), ncol = 1),
  epsilon = 0
)
```

Arguments

alpha	the vectors of parameters $c(a.mu, a.pi, b)$ concatenated
Y	the vector of counts
A.mu	matrix of the model (see Details, default=empty)
B.mu	matrix of the model (see Details, default=empty)
C.mu	matrix of the model (see Details, default=zero)
A.pi	matrix of the model (see Details, default=empty)
B.pi	matrix of the model (see Details, default=empty)
C.pi	matrix of the model (see Details, default=zero)
C.theta	matrix of the model (see Details, default=zero)
epsilon	regularization parameter. A vector of the same length as alpha if each coordinate of alpha has a specific regularization, or just a scalar is the regularization is the same for all coordinates of alpha. Default=0.

Details

The regression model is parametrized as follows:

$$\log(\mu) = A_{\mu} * a_{\mu} + B_{\mu} * b + C_{\mu}$$

$$\text{logit}(\Pi) = A_{\pi} * a_{\pi} + B_{\pi} * b$$

$$\log(\theta) = C_{\theta}$$

where μ, Π, θ are respectively the vector of mean parameters of the NB distribution, the vector of probabilities of the zero component, and the vector of inverse dispersion parameters. Note that the b vector is shared between the mean of the negative binomial and the probability of zero. The log-likelihood of a vector of parameters $\alpha = (a_{\mu}; a_{\pi}; b)$ is penalized by a regularization term $\epsilon ||\alpha||^2/2$ if ϵ is a scalar, or $\sum_i \epsilon_i \alpha_i^2/2$ if ϵ is a vector of the same size as α to allow for differential regularization among the parameters.

Value

the penalized log-likelihood.

```
zinb.loglik.regression.gradient
```

Gradient of the penalized log-likelihood of the ZINB regression model

Description

This function computes the gradient of the penalized log-likelihood of a ZINB regression model given a vector of counts.

Usage

```
zinb.loglik.regression.gradient(
  alpha,
  Y,
  A.mu = matrix(nrow = length(Y), ncol = 0),
  B.mu = matrix(nrow = length(Y), ncol = 0),
  C.mu = matrix(0, nrow = length(Y), ncol = 1),
  A.pi = matrix(nrow = length(Y), ncol = 0),
  B.pi = matrix(nrow = length(Y), ncol = 0),
  C.pi = matrix(0, nrow = length(Y), ncol = 1),
  C.theta = matrix(0, nrow = length(Y), ncol = 1),
  epsilon = 0
)
```

Arguments

<code>alpha</code>	the vectors of parameters $c(a_{\mu}, a_{\pi}, b)$ concatenated
<code>Y</code>	the vector of counts
<code>A.mu</code>	matrix of the model (see Details, default=empty)
<code>B.mu</code>	matrix of the model (see Details, default=empty)
<code>C.mu</code>	matrix of the model (see Details, default=zero)

A.pi	matrix of the model (see Details, default=empty)
B.pi	matrix of the model (see Details, default=empty)
C.pi	matrix of the model (see Details, default=zero)
C.theta	matrix of the model (see Details, default=zero)
epsilon	regularization parameter. A vector of the same length as alpha if each coordinate of alpha has a specific regularization, or just a scalar is the regularization is the same for all coordinates of alpha. Default=0.

Details

The regression model is described in [zinb.loglik.regression](#).

Value

The gradient of the penalized log-likelihood.

See Also

[zinb.loglik.regression](#)

zinb.regression.parseModel

Parse ZINB regression model

Description

Given the parameters of a ZINB regression model, this function parses the model and computes the vector of $\log(\mu)$, $\text{logit}(\pi)$, and the dimensions of the different components of the vector of parameters. See [zinb.loglik.regression](#) for details of the ZINB regression model and its parameters.

Usage

```
zinb.regression.parseModel(alpha, A.mu, B.mu, C.mu, A.pi, B.pi, C.pi)
```

Arguments

alpha	the vectors of parameters $c(a.\mu, a.\pi, b)$ concatenated
A.mu	matrix of the model (see above, default=empty)
B.mu	matrix of the model (see above, default=empty)
C.mu	matrix of the model (see above, default=zero)
A.pi	matrix of the model (see above, default=empty)
B.pi	matrix of the model (see above, default=empty)
C.pi	matrix of the model (see above, default=zero)

Value

A list with slots `logMu`, `logitPi`, `dim.alpha` (a vector of length 3 with the dimension of each of the vectors `a.mu`, `a.pi` and `b` in `alpha`), and `start.alpha` (a vector of length 3 with the starting indices of the 3 vectors in `alpha`)

See Also

[zinb.loglik.regression](#)

zinbAIC	<i>Compute the AIC or BIC of a model given some data</i>
---------	--

Description

Given a statistical model and some data, these functions compute the AIC or BIC of the model given the data, i.e., the AIC/BIC of the data under the model.

Usage

```
zinbAIC(model, x)

zinbBIC(model, x)

## S4 method for signature 'ZinbModel,matrix'
zinbAIC(model, x)

## S4 method for signature 'ZinbModel,matrix'
zinbBIC(model, x)
```

Arguments

model	an object that describes a statistical model.
x	an object that describes data.

Value

the AIC/BIC of the model.

Functions

- `zinbAIC(model = ZinbModel, x = matrix)`: returns the AIC of the ZINB model.
- `zinbBIC(model = ZinbModel, x = matrix)`: returns the BIC of the ZINB model.

Examples

```
se <- SummarizedExperiment(matrix(rpois(60, lambda=5), nrow=10, ncol=6),
  colData = data.frame(bio = gl(2, 3)))
m <- zinbFit(se, X=model.matrix(~bio, data=colData(se)),
  BPPARAM=BiocParallel::SerialParam())
zinbAIC(m, t(assay(se)))
zinbBIC(m, t(assay(se)))
```

`zinbFit`*Fit a ZINB regression model*

Description

Given an object with the data, it fits a ZINB model.

Usage

```
zinbFit(Y, ...)  
  
## S4 method for signature 'SummarizedExperiment'  
zinbFit(  
  Y,  
  X,  
  V,  
  K,  
  which_assay,  
  commondispersion = TRUE,  
  zeroinflation = TRUE,  
  verbose = FALSE,  
  nb.repeat.initialize = 2,  
  maxiter.optimize = 25,  
  stop.epsilon.optimize = 1e-04,  
  BPPARAM = BiocParallel::bpparam(),  
  ...  
)  
  
## S4 method for signature 'matrix'  
zinbFit(  
  Y,  
  X,  
  V,  
  K,  
  commondispersion = TRUE,  
  zeroinflation = TRUE,  
  verbose = FALSE,  
  nb.repeat.initialize = 2,  
  maxiter.optimize = 25,  
  stop.epsilon.optimize = 1e-04,  
  BPPARAM = BiocParallel::bpparam(),  
  ...  
)  
  
## S4 method for signature 'dgCMatrix'  
zinbFit(Y, ...)
```

Arguments

<code>Y</code>	The data (genes in rows, samples in columns).
<code>...</code>	Additional parameters to describe the model, see zinbModel .

<code>X</code>	The design matrix containing sample-level covariates, one sample per row. If missing, <code>X</code> will contain only an intercept. If <code>Y</code> is a <code>SummarizedExperiment</code> object, <code>X</code> can be a formula using the variables in the <code>colData</code> slot of <code>Y</code> .
<code>V</code>	The design matrix containing gene-level covariates, one gene per row. If missing, <code>V</code> will contain only an intercept. If <code>Y</code> is a <code>SummarizedExperiment</code> object, <code>V</code> can be a formula using the variables in the <code>rowData</code> slot of <code>Y</code> .
<code>K</code>	integer. Number of latent factors.
<code>which_assay</code>	numeric or character. Which assay of <code>Y</code> to use (only if <code>Y</code> is a <code>SummarizedExperiment</code>).
<code>commondispersion</code>	Whether or not a single dispersion for all features is estimated (default <code>TRUE</code>).
<code>zeroinflation</code>	Whether or not a ZINB model should be fitted. If <code>FALSE</code> , a negative binomial model is fitted instead.
<code>verbose</code>	Print helpful messages.
<code>nb.repeat.initialize</code>	Number of iterations for the initialization of <code>beta_mu</code> and <code>gamma_mu</code> .
<code>maxiter.optimize</code>	maximum number of iterations for the optimization step (default 25).
<code>stop.epsilon.optimize</code>	stopping criterion in the optimization step, when the relative gain in likelihood is below <code>epsilon</code> (default 0.0001).
<code>BPPARAM</code>	object of class <code>bpparamClass</code> that specifies the back-end to be used for computations. See bpparam for details.

Details

By default, i.e., if no arguments other than `Y` are passed, the model is fitted with an intercept for the regression across-samples and one intercept for the regression across genes, both for `mu` and for `pi`.

This means that by default the model is fitted with $X_{\text{mu}} = X_{\text{pi}} = 1_n$ and $V_{\text{mu}} = V_{\text{pi}} = 1_J$. If the user explicitly passes the design matrices, this behavior is overwritten, i.e., the user needs to explicitly include the intercept in the design matrices.

If `Y` is a `SummarizedExperiment`, the function uses the assay named "counts", if any, or the first assay.

Currently, if `Y` is a `sparseMatrix`, this calls the `zinbFit` method on `as.matrix(Y)`

Value

An object of class `ZinbModel` that has been fitted by penalized maximum likelihood on the data.

Methods (by class)

- `zinbFit(SummarizedExperiment)`: `Y` is a [SummarizedExperiment](#).
- `zinbFit(matrix)`: `Y` is a matrix of counts (genes in rows).
- `zinbFit(dgCMatrix)`: `Y` is a sparse matrix of counts (genes in rows).

See Also

[model.matrix](#).

Examples

```
se <- SummarizedExperiment(matrix(rpois(60, lambda=5), nrow=10, ncol=6),
                              colData = data.frame(bio = gl(2, 3)))

m <- zinbFit(se, X=model.matrix(~bio, data=colData(se)),
             BPPARAM=BiocParallel::SerialParam())
bio <- gl(2, 3)
m <- zinbFit(matrix(rpois(60, lambda=5), nrow=10, ncol=6),
             X=model.matrix(~bio), BPPARAM=BiocParallel::SerialParam())
```

zinbInitialize

*Initialize the parameters of a ZINB regression model***Description**

The initialization performs quick optimization of the parameters with several simplifying assumptions compared to the true model: non-zero counts are modeled as log-Gaussian, zeros are modeled as dropouts. The dispersion parameter is not modified.

Usage

```
zinbInitialize(
  m,
  Y,
  nb.repeat = 2,
  it.max = 100,
  BPPARAM = BiocParallel::bpparam()
)
```

Arguments

<code>m</code>	The model of class <code>ZinbModel</code>
<code>Y</code>	The matrix of counts.
<code>nb.repeat</code>	Number of iterations for the estimation of <code>beta_mu</code> and <code>gamma_mu</code> .
<code>it.max</code>	Maximum number of iterations in <code>softImpute</code> .
<code>BPPARAM</code>	object of class <code>bpparamClass</code> that specifies the back-end to be used for computations. See bpparam for details.

Value

An object of class `ZinbModel` similar to the one given as argument with modified parameters `alpha_mu`, `alpha_pi`, `beta_mu`, `beta_pi`, `gamma_mu`, `gamma_pi`, `W`.

Examples

```
Y <- matrix(rpois(60, lambda=2), 6, 10)
bio <- gl(2, 3)
time <- rnorm(6)
gc <- rnorm(10)
m <- zinbModel(Y, X=model.matrix(~bio + time), V=model.matrix(~gc),
              which_X_pi=1L, which_V_mu=1L, K=1)
m <- zinbInitialize(m, Y, BPPARAM=BiocParallel::SerialParam())
```

zinbModel	<i>Initialize an object of class ZinbModel</i>
-----------	--

Description

Initialize an object of class ZinbModel

Usage

```
zinbModel(
  X,
  V,
  O_mu,
  O_pi,
  which_X_mu,
  which_X_pi,
  which_V_mu,
  which_V_pi,
  W,
  beta_mu,
  beta_pi,
  gamma_mu,
  gamma_pi,
  alpha_mu,
  alpha_pi,
  zeta,
  epsilon,
  epsilon_beta_mu,
  epsilon_gamma_mu,
  epsilon_beta_pi,
  epsilon_gamma_pi,
  epsilon_W,
  epsilon_alpha,
  epsilon_zeta,
  epsilon_min_logit,
  n,
  J,
  K
)
```

Arguments

X	matrix. The design matrix containing sample-level covariates, one sample per row.
V	matrix. The design matrix containing gene-level covariates, one gene per row.
O_mu	matrix. The offset matrix for mu.
O_pi	matrix. The offset matrix for pi.
which_X_mu	integer. Indices of which columns of X to use in the regression of mu.
which_X_pi	integer. Indices of which columns of X to use in the regression of pi.

<code>which_V_mu</code>	integer. Indices of which columns of <code>V</code> to use in the regression of <code>mu</code> .
<code>which_V_pi</code>	integer. Indices of which columns of <code>V</code> to use in the regression of <code>pi</code> .
<code>W</code>	matrix. The factors of sample-level latent factors.
<code>beta_mu</code>	matrix or NULL. The coefficients of <code>X</code> in the regression of <code>mu</code> .
<code>beta_pi</code>	matrix or NULL. The coefficients of <code>X</code> in the regression of <code>pi</code> .
<code>gamma_mu</code>	matrix or NULL. The coefficients of <code>V</code> in the regression of <code>mu</code> .
<code>gamma_pi</code>	matrix or NULL. The coefficients of <code>V</code> in the regression of <code>pi</code> .
<code>alpha_mu</code>	matrix or NULL. The coefficients of <code>W</code> in the regression of <code>mu</code> .
<code>alpha_pi</code>	matrix or NULL. The coefficients of <code>W</code> in the regression of <code>pi</code> .
<code>zeta</code>	numeric. A vector of log of inverse dispersion parameters.
<code>epsilon</code>	nonnegative scalar. Regularization parameter.
<code>epsilon_beta_mu</code>	nonnegative scalar. Regularization parameter for <code>beta_mu</code> .
<code>epsilon_gamma_mu</code>	nonnegative scalar. Regularization parameter for <code>gamma_mu</code> .
<code>epsilon_beta_pi</code>	nonnegative scalar. Regularization parameter for <code>beta_pi</code> .
<code>epsilon_gamma_pi</code>	nonnegative scalar. Regularization parameter for <code>gamma_pi</code> .
<code>epsilon_W</code>	nonnegative scalar. Regularization parameter for <code>W</code> .
<code>epsilon_alpha</code>	nonnegative scalar. Regularization parameter for <code>alpha</code> (both <code>alpha_mu</code> and <code>alpha_pi</code>).
<code>epsilon_zeta</code>	nonnegative scalar. Regularization parameter for <code>zeta</code> .
<code>epsilon_min_logit</code>	scalar. Minimum regularization parameter for parameters of the logit model, including the intercept.
<code>n</code>	integer. Number of samples.
<code>J</code>	integer. Number of genes.
<code>K</code>	integer. Number of latent factors.

Details

This is a wrapper around the `new()` function to create an instance of class `ZinbModel`. Rarely, the user will need to create a `ZinbModel` object from scratch, as typically this is the result of [zinbFit](#).

If any of `X`, `V`, `W` matrices are passed, `n`, `J`, and `K` are inferred. Alternatively, the user can specify one or more of `n`, `J`, and `K`.

The regularization parameters can be set by a unique parameter `epsilon` or specific values for the different regularization parameters can also be provided. If only `epsilon` is specified, the other parameters take the following values:

- `epsilon_beta = epsilon/J`
- `epsilon_gamma = epsilon/n`
- `epsilon_W = epsilon/n`
- `epsilon_alpha = epsilon/J`
- `epsilon_zeta = epsilon`

We empirically found that large values of epsilon provide a more stable estimation of W .

A call with no argument has the following default values: $n = 50$, $J = 100$, $K = 0$, $\text{epsilon} = J$.

Although it is possible to create new instances of the class by calling this function, this is not the most common way of creating ZinbModel objects. The main use of the class is within the [zinbFit](#) function.

Value

an object of class [ZinbModel](#).

Examples

```
a <- zinbModel()
nSamples(a)
nFeatures(a)
nFactors(a)
nParams(a)
```

ZinbModel-class	<i>Class ZinbModel</i>
-----------------	------------------------

Description

Objects of this class store all the values needed to work with a zero-inflated negative binomial (ZINB) model, as described in the vignette. They contain all information to fit a model by penalized maximum likelihood or simulate data from a model.

Usage

```
## S4 method for signature 'ZinbModel'
show(object)

## S4 method for signature 'ZinbModel'
nSamples(x)

## S4 method for signature 'ZinbModel'
nFeatures(x)

## S4 method for signature 'ZinbModel'
nFactors(x)

## S4 method for signature 'ZinbModel'
getX_mu(object, intercept = TRUE)

## S4 method for signature 'ZinbModel'
getX_pi(object, intercept = TRUE)

## S4 method for signature 'ZinbModel'
getV_mu(object, intercept = TRUE)

## S4 method for signature 'ZinbModel'
```

```
getV_pi(object, intercept = TRUE)

## S4 method for signature 'ZinbModel'
getLogMu(object)

## S4 method for signature 'ZinbModel'
getMu(object)

## S4 method for signature 'ZinbModel'
getLogitPi(object)

## S4 method for signature 'ZinbModel'
getPi(object)

## S4 method for signature 'ZinbModel'
getZeta(object)

## S4 method for signature 'ZinbModel'
getPhi(object)

## S4 method for signature 'ZinbModel'
getTheta(object)

## S4 method for signature 'ZinbModel'
getEpsilon_beta_mu(object)

## S4 method for signature 'ZinbModel'
getEpsilon_gamma_mu(object)

## S4 method for signature 'ZinbModel'
getEpsilon_beta_pi(object)

## S4 method for signature 'ZinbModel'
getEpsilon_gamma_pi(object)

## S4 method for signature 'ZinbModel'
getEpsilon_W(object)

## S4 method for signature 'ZinbModel'
getEpsilon_alpha(object)

## S4 method for signature 'ZinbModel'
getEpsilon_zeta(object)

## S4 method for signature 'ZinbModel'
getW(object)

## S4 method for signature 'ZinbModel'
getBeta_mu(object)

## S4 method for signature 'ZinbModel'
getBeta_pi(object)
```



```
## S4 method for signature 'ZinbModel'
getGamma_mu(object)

## S4 method for signature 'ZinbModel'
getGamma_pi(object)

## S4 method for signature 'ZinbModel'
getAlpha_mu(object)

## S4 method for signature 'ZinbModel'
getAlpha_pi(object)
```

Arguments

object	an object of class ZinbModel.
x	an object of class ZinbModel.
intercept	logical. Whether to return the intercept (ignored if the design matrix has no intercept). Default TRUE

Details

For the full description of the model see the model vignette. Internally, the slots are checked so that the matrices are of the appropriate dimensions: in particular, X , O_{μ} , O_{π} , and W need to have n rows, V needs to have J rows, ζ must be of length J .

Value

`nSamples` returns the number of samples; `nFeatures` returns the number of features; `nFactors` returns the number of latent factors.

Methods (by generic)

- `show(ZinbModel)`: show useful info on the object.
- `nSamples(ZinbModel)`: returns the number of samples.
- `nFeatures(ZinbModel)`: returns the number of features.
- `nFactors(ZinbModel)`: returns the number of latent factors.
- `getX_mu(ZinbModel)`: returns the sample-level design matrix for μ .
- `getX_pi(ZinbModel)`: returns the sample-level design matrix for π .
- `getV_mu(ZinbModel)`: returns the gene-level design matrix for μ .
- `getV_pi(ZinbModel)`: returns the sample-level design matrix for π .
- `getLogMu(ZinbModel)`: returns the logarithm of the mean of the non-zero component.
- `getMu(ZinbModel)`: returns the mean of the non-zero component.
- `getLogitPi(ZinbModel)`: returns the logit-probability of zero.
- `getPi(ZinbModel)`: returns the probability of zero.
- `getZeta(ZinbModel)`: returns the log of the inverse of the dispersion parameter.
- `getPhi(ZinbModel)`: returns the dispersion parameter.
- `getTheta(ZinbModel)`: returns the inverse of the dispersion parameter.

- `getEpsilon_beta_mu(ZinbModel)`: returns the regularization parameters for `beta_mu`.
- `getEpsilon_gamma_mu(ZinbModel)`: returns the regularization parameters for `gamma_mu`.
- `getEpsilon_beta_pi(ZinbModel)`: returns the regularization parameters for `beta_pi`.
- `getEpsilon_gamma_pi(ZinbModel)`: returns the regularization parameters for `gamma_pi`.
- `getEpsilon_W(ZinbModel)`: returns the regularization parameters for `W`.
- `getEpsilon_alpha(ZinbModel)`: returns the regularization parameters for `alpha`.
- `getEpsilon_zeta(ZinbModel)`: returns the regularization parameters for `zeta`.
- `getW(ZinbModel)`: returns the matrix `W` of inferred sample-level covariates.
- `getBeta_mu(ZinbModel)`: returns the matrix `beta_mu` of inferred parameters.
- `getBeta_pi(ZinbModel)`: returns the matrix `beta_pi` of inferred parameters.
- `getGamma_mu(ZinbModel)`: returns the matrix `gamma_mu` of inferred parameters.
- `getGamma_pi(ZinbModel)`: returns the matrix `gamma_pi` of inferred parameters.
- `getAlpha_mu(ZinbModel)`: returns the matrix `alpha_mu` of inferred parameters.
- `getAlpha_pi(ZinbModel)`: returns the matrix `alpha_pi` of inferred parameters.

Slots

`X` matrix. The design matrix containing sample-level covariates, one sample per row.

`V` matrix. The design matrix containing gene-level covariates, one gene per row.

`O_mu` matrix. The offset matrix for `mu`.

`O_pi` matrix. The offset matrix for `pi`.

`which_X_mu` integer. Indices of which columns of `X` to use in the regression of `mu`.

`which_V_mu` integer. Indices of which columns of `V` to use in the regression of `mu`.

`which_X_pi` integer. Indices of which columns of `X` to use in the regression of `pi`.

`which_V_pi` integer. Indices of which columns of `V` to use in the regression of `pi`.

`X_mu_intercept` logical. TRUE if `X_mu` contains an intercept.

`X_pi_intercept` logical. TRUE if `X_pi` contains an intercept.

`V_mu_intercept` logical. TRUE if `V_mu` contains an intercept.

`V_pi_intercept` logical. TRUE if `V_pi` contains an intercept.

`W` matrix. The factors of sample-level latent factors.

`beta_mu` matrix or NULL. The coefficients of `X` in the regression of `mu`.

`gamma_mu` matrix or NULL. The coefficients of `V` in the regression of `mu`.

`alpha_mu` matrix or NULL. The coefficients of `W` in the regression of `mu`.

`beta_pi` matrix or NULL. The coefficients of `X` in the regression of `pi`.

`gamma_pi` matrix or NULL. The coefficients of `V` in the regression of `pi`.

`alpha_pi` matrix or NULL. The coefficients of `W` in the regression of `pi`.

`zeta` numeric. A vector of log of inverse dispersion parameters.

`epsilon_beta_mu` nonnegative scalar. Regularization parameter for `beta_mu`

`epsilon_gamma_mu` nonnegative scalar. Regularization parameter for `gamma_mu`

`epsilon_beta_pi` nonnegative scalar. Regularization parameter for `beta_pi`

`epsilon_gamma_pi` nonnegative scalar. Regularization parameter for `gamma_pi`

epsilon_W nonnegative scalar. Regularization parameter for W

epsilon_alpha nonnegative scalar. Regularization parameter for alpha (both alpha_mu and alpha_pi)

epsilon_zeta nonnegative scalar. Regularization parameter for zeta

epsilon_min_logit scalar. Minimum regularization parameter for parameters of the logit model, including the intercept.

zinbOptimize

Optimize the parameters of a ZINB regression model

Description

The parameters of the model given as argument are optimized by penalized maximum likelihood on the count matrix given as argument. It is recommended to call `zinb_initialize` before this function to have good starting point for optimization, since the optimization problem is not convex and can only converge to a local minimum.

Usage

```
zinbOptimize(
  m,
  Y,
  comondispersion = TRUE,
  maxiter = 25,
  stop.epsilon = 1e-04,
  verbose = FALSE,
  BPPARAM = BiocParallel::bpparam()
)
```

Arguments

m	The model of class <code>ZinbModel</code>
Y	The matrix of counts.
comondispersion	Whether the dispersion is the same for all features (default=TRUE)
maxiter	maximum number of iterations (default 25)
stop.epsilon	stopping criterion, when the relative gain in likelihood is below epsilon (default 0.0001)
verbose	print information (default FALSE)
BPPARAM	object of class <code>bpparamClass</code> that specifies the back-end to be used for computations. See bpparam for details.

Value

An object of class `ZinbModel` similar to the one given as argument with modified parameters `alpha_mu`, `alpha_pi`, `beta_mu`, `beta_pi`, `gamma_mu`, `gamma_pi`, `W`.

Examples

```
Y = matrix(10, 3, 5)
m = zinbModel(n=NROW(Y), J=NCOL(Y))
m = zinbInitialize(m, Y, BPPARAM=BiocParallel::SerialParam())
m = zinbOptimize(m, Y, BPPARAM=BiocParallel::SerialParam())
```

zinbOptimizeDispersion

Optimize the dispersion parameters of a ZINB regression model

Description

The dispersion parameters of the model are optimized by penalized maximum likelihood on the count matrix given as argument.

Usage

```
zinbOptimizeDispersion(
  J,
  mu,
  logitPi,
  epsilon,
  Y,
  commondispersion = TRUE,
  BPPARAM = BiocParallel::bpparam()
)
```

Arguments

J	The number of genes.
mu	the matrix containing the mean of the negative binomial.
logitPi	the matrix containing the logit of the probability parameter of the zero-inflation part of the model.
epsilon	the regularization parameter.
Y	The matrix of counts.
commondispersion	Whether or not a single dispersion for all features is estimated (default TRUE)
BPPARAM	object of class <code>bpparamClass</code> that specifies the back-end to be used for computations. See bpparam for details.

Value

An object of class `ZinbModel` similar to the one given as argument with modified parameters `zeta`.

Examples

```
Y = matrix(10, 3, 5)
m = zinbModel(n=NROW(Y), J=NCOL(Y))
m = zinbInitialize(m, Y, BPPARAM=BiocParallel::SerialParam())
m = zinbOptimizeDispersion(NROW(Y), getMu(m), getLogitPi(m),
  getEpsilon_zeta(m), Y, BPPARAM=BiocParallel::SerialParam())
```

zinbSim

Simulate counts from a zero-inflated negative binomial model

Description

Given an object that describes zero-inflated negative binomial distribution, simulate counts from the distribution.

Usage

```
zinbSim(object, seed, ...)

## S4 method for signature 'ZinbModel'
zinbSim(object, seed)
```

Arguments

object	an object that describes a matrix of zero-inflated negative binomial.
seed	an optional integer to specify how the random number generator should be initialized with a call to <code>set.seed</code> . If missing, the random generator state is not changed.
...	additional arguments.

Value

A list with the following elements.

- `counts` the matrix with the simulated counts.
- `dataNB` the data simulated from the negative binomial.
- `dataDropout` the data simulated from the binomial process.
- `zeroFraction` the fraction of zeros.

Methods (by class)

- `zinbSim(ZinbModel)`: simulate from a ZINB distribution.

Examples

```
a <- zinbModel(n=5, J=10)
zinbSim(a)
```

zinbsurf	<i>Perform dimensionality reduction using a ZINB regression model for large datasets.</i>
----------	---

Description

Given an object with the data, it performs dimensionality reduction using a ZINB regression model with gene and cell-level covariates on a random subset of the data. It then projects the remaining data onto the lower dimensional space.

Usage

```
zinbsurf(Y, ...)

## S4 method for signature 'SummarizedExperiment'
zinbsurf(
  Y,
  X,
  V,
  K,
  which_assay,
  which_genes,
  zeroinflation = TRUE,
  prop_fit = 0.1,
  BPPARAM = BiocParallel::bpparam(),
  verbose = FALSE,
  ...
)
```

Arguments

Y	The data (genes in rows, samples in columns). Currently implemented only for SummarizedExperiment.
...	Additional parameters to describe the model, see zinbModel .
X	The design matrix containing sample-level covariates, one sample per row. If missing, X will contain only an intercept. If Y is a SummarizedExperiment object, X can be a formula using the variables in the colData slot of Y.
V	The design matrix containing gene-level covariates, one gene per row. If missing, V will contain only an intercept. If Y is a SummarizedExperiment object, V can be a formula using the variables in the rowData slot of Y.
K	integer. Number of latent factors. Specify $K = 0$ if only computing observational weights.
which_assay	numeric or character. Which assay of Y to use. If missing, if 'assayNames(Y)' contains "counts" then that is used. Otherwise, the first assay is used.
which_genes	character. Which genes to use to estimate W (see details). Ignored if fitted_model is provided.
zeroinflation	Whether or not a ZINB model should be fitted. If FALSE, a negative binomial model is fitted instead.
prop_fit	numeric between 0 and 1. The proportion of cells to use for the zinbwave fit.

BPPARAM	object of class <code>bpparamClass</code> that specifies the back-end to be used for computations. See bpparam for details.
verbose	Print helpful messages.

Details

This function implements an approximate strategy, in which the full zinbwave model is fit only on a random subset of the data (controlled by the `prop_fit` parameter). The rest of the samples are subsequently projected onto the low-rank space. This strategy is much faster and uses less memory than the full [zinbwave](#) method. It is recommended with extremely large datasets.

By default `zinbsurf` uses all genes to estimate W . However, we recommend to use the top 1,000 most variable genes for this step. In general, a user can specify any custom set of genes to be used to estimate W , by specifying either a vector of gene names, or a single character string corresponding to a column of the `rowData`.

Value

An object of class `SingleCellExperiment`; the dimensionality reduced matrix is stored in the `reducedDims` slot.

Methods (by class)

- `zinbsurf(SummarizedExperiment)`: Y is a [SummarizedExperiment](#).

Examples

```
se <- SingleCellExperiment(assays = list(counts = matrix(rpois(60, lambda=5),
                                                         nrow=10, ncol=6)),
                           colData = data.frame(bio = gl(2, 3)))
colnames(se) <- paste0("sample", 1:6)
m <- zinbsurf(se, X="~bio", K = 1, prop_fit = .5, which_assay = 1,
              BPPARAM=BiocParallel::SerialParam())
```

zinbwave	<i>Perform dimensionality reduction using a ZINB regression model with gene and cell-level covariates.</i>
----------	--

Description

Given an object with the data, it performs dimensionality reduction using a ZINB regression model with gene and cell-level covariates.

Usage

```
zinbwave(Y, ...)

## S4 method for signature 'SummarizedExperiment'
zinbwave(
  Y,
  X,
  V,
  K = 2,
```

```

    fitted_model,
    which_assay,
    which_genes,
    commondispersion = TRUE,
    zeroinflation = TRUE,
    verbose = FALSE,
    nb.repeat.initialize = 2,
    maxiter.optimize = 25,
    stop.epsilon.optimize = 1e-04,
    BPPARAM = BiocParallel::bpparam(),
    normalizedValues = FALSE,
    residuals = FALSE,
    imputedValues = FALSE,
    observationalWeights = FALSE,
    ...
)

```

Arguments

Y	The data (genes in rows, samples in columns). Currently implemented only for SummarizedExperiment.
...	Additional parameters to describe the model, see zinbModel .
X	The design matrix containing sample-level covariates, one sample per row. If missing, X will contain only an intercept. If Y is a SummarizedExperiment object, X can be a formula using the variables in the colData slot of Y.
V	The design matrix containing gene-level covariates, one gene per row. If missing, V will contain only an intercept. If Y is a SummarizedExperiment object, V can be a formula using the variables in the rowData slot of Y.
K	integer. Number of latent factors. Specify $K = 0$ if only computing observational weights.
fitted_model	a ZinbModel object.
which_assay	numeric or character. Which assay of Y to use. If missing, if 'assayNames(Y)' contains "counts" then that is used. Otherwise, the first assay is used.
which_genes	character. Which genes to use to estimate W (see details). Ignored if fitted_model is provided.
commondispersion	Whether or not a single dispersion for all features is estimated (default TRUE).
zeroinflation	Whether or not a ZINB model should be fitted. If FALSE, a negative binomial model is fitted instead.
verbose	Print helpful messages.
nb.repeat.initialize	Number of iterations for the initialization of beta_mu and gamma_mu.
maxiter.optimize	maximum number of iterations for the optimization step (default 25).
stop.epsilon.optimize	stopping criterion in the optimization step, when the relative gain in likelihood is below epsilon (default 0.0001).
BPPARAM	object of class bpparamClass that specifies the back-end to be used for computations. See bpparam for details.

<code>normalizedValues</code>	indicates whether or not you want to compute normalized values for the counts after adjusting for gene and cell-level covariates.
<code>residuals</code>	indicates whether or not you want to compute the residuals of the ZINB model. Deviance residuals are computed.
<code>imputedValues</code>	indicates whether or not you want to compute the imputed counts of the ZINB model.
<code>observationalWeights</code>	indicates whether to compute the observational weights for differential expression (see vignette).

Details

For visualization (heatmaps, ...), please use the normalized values. It corresponds to the deviance residuals when the W is not included in the model but the gene and cell-level covariates are. As a result, when W is not included in the model, the deviance residuals should capture the biology. Note that we do not recommend to use the normalized values for any downstream analysis (such as clustering, or differential expression), but only for visualization.

If one has already fitted a model using `ZinbModel`, the object containing such model can be used as input of `zinbwave` to save the resulting W into a `SummarizedExperiment` and optionally compute residuals and normalized values, without the need for re-fitting the model.

By default `zinbwave` uses all genes to estimate W . However, we recommend to use the top 1,000 most variable genes for this step. In general, a user can specify any custom set of genes to be used to estimate W , by specifying either a vector of gene names, or a single character string corresponding to a column of the `rowData`.

Note that if both `which_genes` is specified and at least one among `observationalWeights`, `imputedValues`, `residuals`, and `normalizedValues` is `TRUE`, the model needs to be fit twice.

Value

An object of class `SingleCellExperiment`; the dimensionality reduced matrix is stored in the `reducedDims` slot and optionally normalized values and residuals are added in the list of assays.

Methods (by class)

- `zinbwave(SummarizedExperiment)`: Y is a `SummarizedExperiment`.

Examples

```
se <- SingleCellExperiment(assays = list(counts = matrix(rpois(60, lambda=5),
                                                         nrow=10, ncol=6)),
                           colData = data.frame(bio = gl(2, 3)))

m <- zinbwave(se, X="~bio", BPPARAM=BiocParallel::SerialParam())
```

Index

bpparam, [35](#), [36](#), [43](#), [44](#), [47](#), [48](#)

computeDevianceResiduals, [3](#)

computeObservationalWeights, [4](#)

data.frame, [20](#)

getAlpha_mu, [4](#)

getAlpha_mu, ZinbModel-method
(ZinbModel-class), [39](#)

getAlpha_pi, [5](#)

getAlpha_pi, ZinbModel-method
(ZinbModel-class), [39](#)

getBeta_mu, [5](#)

getBeta_mu, ZinbModel-method
(ZinbModel-class), [39](#)

getBeta_pi, [6](#)

getBeta_pi, ZinbModel-method
(ZinbModel-class), [39](#)

getEpsilon_alpha, [6](#)

getEpsilon_alpha, ZinbModel-method
(ZinbModel-class), [39](#)

getEpsilon_beta_mu, [7](#)

getEpsilon_beta_mu, ZinbModel-method
(ZinbModel-class), [39](#)

getEpsilon_beta_pi, [7](#)

getEpsilon_beta_pi, ZinbModel-method
(ZinbModel-class), [39](#)

getEpsilon_gamma_mu, [8](#)

getEpsilon_gamma_mu, ZinbModel-method
(ZinbModel-class), [39](#)

getEpsilon_gamma_pi, [8](#)

getEpsilon_gamma_pi, ZinbModel-method
(ZinbModel-class), [39](#)

getEpsilon_W, [9](#)

getEpsilon_W, ZinbModel-method
(ZinbModel-class), [39](#)

getEpsilon_zeta, [9](#)

getEpsilon_zeta, ZinbModel-method
(ZinbModel-class), [39](#)

getGamma_mu, [10](#)

getGamma_mu, ZinbModel-method
(ZinbModel-class), [39](#)

getGamma_pi, [10](#)

getGamma_pi, ZinbModel-method
(ZinbModel-class), [39](#)

getLogitPi, [11](#)

getLogitPi, ZinbModel-method
(ZinbModel-class), [39](#)

getLogMu, [12](#)

getLogMu, ZinbModel-method
(ZinbModel-class), [39](#)

getMu, [12](#)

getMu, ZinbModel-method
(ZinbModel-class), [39](#)

getPhi, [13](#)

getPhi, ZinbModel-method
(ZinbModel-class), [39](#)

getPi, [13](#)

getPi, ZinbModel-method
(ZinbModel-class), [39](#)

getTheta, [14](#)

getTheta, ZinbModel-method
(ZinbModel-class), [39](#)

getV_mu, [15](#)

getV_mu, ZinbModel-method
(ZinbModel-class), [39](#)

getV_pi, [15](#)

getV_pi, ZinbModel-method
(ZinbModel-class), [39](#)

getW, [16](#)

getW, ZinbModel-method
(ZinbModel-class), [39](#)

getX_mu, [16](#)

getX_mu, ZinbModel-method
(ZinbModel-class), [39](#)

getX_pi, [17](#)

getX_pi, ZinbModel-method
(ZinbModel-class), [39](#)

getZeta, [17](#)

getZeta, ZinbModel-method
(ZinbModel-class), [39](#)

glmFit, [18](#)

glmLRT, [18–20](#)

glmWeightedF, [18](#)

imputeZeros, [19](#)

independentFiltering, [20](#)

loglik, [20](#)
 loglik, ZinbModel, matrix-method
 (loglik), [20](#)

 model.matrix, [35](#)

 nFactors, [21](#)
 nFactors, ZinbModel-method
 (ZinbModel-class), [39](#)
 nFeatures, [22](#)
 nFeatures, ZinbModel-method
 (ZinbModel-class), [39](#)
 nParams, [22](#)
 nParams, ZinbModel-method (nParams), [22](#)
 nSamples, [23](#)
 nSamples, ZinbModel-method
 (ZinbModel-class), [39](#)

 orthogonalizeTraceNorm, [23](#)

 penalty, [24](#)
 penalty, ZinbModel-method (penalty), [24](#)
 pvalueAdjustment, [25](#)

 results, [20](#)

 show, ZinbModel-method
 (ZinbModel-class), [39](#)
 solveRidgeRegression, [26](#)
 SummarizedExperiment, [35](#), [47](#), [49](#)

 toydata, [27](#)

 zinb.loglik, [27](#), [28](#), [29](#)
 zinb.loglik.dispersion, [28](#), [29](#)
 zinb.loglik.dispersion.gradient, [29](#)
 zinb.loglik.matrix, [29](#)
 zinb.loglik.regression, [30](#), [32](#), [33](#)
 zinb.loglik.regression.gradient, [31](#)
 zinb.regression.parseModel, [32](#)
 zinbAIC, [33](#)
 zinbAIC, ZinbModel, matrix-method
 (zinbAIC), [33](#)
 zinbBIC (zinbAIC), [33](#)
 zinbBIC, ZinbModel, matrix-method
 (zinbAIC), [33](#)
 zinbFit, [11](#), [12](#), [14](#), [16](#), [34](#), [38](#), [39](#)
 zinbFit, dgCMatrx-method (zinbFit), [34](#)
 zinbFit, matrix-method (zinbFit), [34](#)
 zinbFit, SummarizedExperiment-method
 (zinbFit), [34](#)
 zinbInitialize, [36](#)
 ZinbModel, [16](#), [39](#), [48](#), [49](#)
 ZinbModel (ZinbModel-class), [39](#)

 zinbModel, [34](#), [37](#), [46](#), [48](#)
 ZinbModel-class, [39](#)
 zinbOptimize, [43](#)
 zinbOptimizeDispersion, [44](#)
 zinbSim, [45](#)
 zinbSim, ZinbModel-method (zinbSim), [45](#)
 zinbsurf, [46](#)
 zinbsurf, SummarizedExperiment-method
 (zinbsurf), [46](#)
 zinbwave, [47](#), [47](#)
 zinbwave, SummarizedExperiment-method
 (zinbwave), [47](#)