Package 'strandCheckR'

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Type Package

Title Calculate strandness information of a bam file

```
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Description This package aims to quantify and remove putative double strand DNA from a strand-
      specific RNA sample. There are also options and methods to plot the positive/negative propor-
      tions of all sliding windows, which allow users to have an idea of how much the sample was con-
      taminated and the appropriate threshold to be used for filtering.
URL https://github.com/UofABioinformaticsHub/strandCheckR
BugReports https://github.com/UofABioinformaticsHub/strandCheckR/issues
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```

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Description

- This package contains functions for checking the strandedness of alignments such as [getStrandFromBamFile()] - The results can easily be visualised using [plotHist()] - Any suspected genomic DNA can also be removed using [filterDNA()], with thresholds for filtering able to be determined manually using [plotWin()]

Author(s)

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See Also

Useful links:

- https://github.com/UofABioinformaticsHub/strandCheckR
- Report bugs at https://github.com/UofABioinformaticsHub/strandCheckR/issues

.calculateStrandCoverage

Calculate the strand information based on coverage

Description

Calculate the coverage coming from '+'/'-' reads in all sliding windows

Usage

```
.calculateStrandCoverage(
  winPosAlignments,
  winNegAlignments,
  winWidth = 1000L,
  winStep = 100L
)
```

Arguments

winPosAlignments

a list that has a 'Coverage' field containing coverage coming from positive reads winNegAlignments

a list that has a 'Coverage' field containing coverage coming from negative reads

winWidth the length of the sliding window, 1000 by default.
winStep the step length to sliding the window, 100 by default.

Value

a list of two vectors, containing a positive/negative coverage of the input positive/negative windows

.calculateStrandNbReads

Calculate the strand information based on the number of reads

Description

Calculate the number of reads coming from '+'/'-' strands in all sliding wndows

Usage

```
. \verb|calculateStrandNbReads(winPosAlignments, winNegAlignments)|\\
```

Arguments

```
winPosAlignments
```

a list that has a 'Win' field that contains information of sliding windows overalapping positive reads

winNegAlignments

a a list that has a 'Win' field that contains information of sliding windows overalapping negative reads 4 .getWinInSequence

Value

a list of two vectors, containing a positive/negative number of reads of the input positive/negative windows

.concatenateAlignments

Concatenate a list of Alignments into One

Description

Concatenate a list of Alignments from multiple sequences into a single object

Usage

.concatenateAlignments(readInfo, seqInfo)

Arguments

readInfo a list returned by scanBam function, each element correspond to a sequence, con-

taining the information of strand, starting position, cigar string, and eventually

flag, qname

seqInfo a data frame that contains some key information of the sequences

Details

This method take a list of alignments across one or more sequences as output by scanBam and concatenates them into a single set of alignments which may include multiple sequences

Value

the concatenated alignments of the input list

.getWinInSequence

Get window data frame with the correct sequence name and position

Description

Get the correct sequence name and position for each window

Usage

```
.getWinInSequence(Win, seqInfo, winWidth = 1000L, winStep = 100L)
```

Arguments

Win a data frame contains the strand information of every window seqInfo a data frame that contains some key information of the sequences

winWidth the length of sliding window

winStep the step length to sliding the window

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Value

A subset of the input object

.keptProbaWin

Get the probability of being kept for each window

Description

Calculate the keeping probability of each window based on its positive/negative proportion

Usage

```
.keptProbaWin(
   winPosAlignments,
   winNegAlignments,
   winWidth,
   winStep,
   threshold,
   pvalueThreshold,
   errorRate,
   mustKeepWin,
   minCov,
   maxCov,
   getWin,
   useCoverage = FALSE
)
```

Arguments

winPosAlignments

an object returned by getWinOverlapEachReadFragment for positive reads

winNegAlignments

an object returned by getWinOverlapEachReadFragment for negative reads

winWidth the width of the sliding window, 1000 by default.

winStep the winStep length to sliding the window, 100 by default.

threshold the strand proportion threshold to test whether to keep a window or not.

pvalueThreshold

threshold of p-value

mustKeepWin the windows that must be kept regardless their strand proportion

minCov In the case that useCoverage=FALSE, if a window has less than minCov reads,

then it will be rejected regardless of the strand proportion. For the case that useCoverage=TRUE, if a window has max coverage least than minCov, then it

will be rejected. 0 by default

maxCov In the case that useCoverage=FALSE, if a window has more than maxCov reads,

then it will be kept regardless of the strand proportion. For the case that useCoverage=TRUE,

if a window has max coverage more than maxCov, then it will be kept. If 0 then

it doesn't have effect on selecting window. 0 by default.

getWin if TRUE, the function will return a data frame containing the information of all

windows. It's FALSE by default.

useCoverage if TRUE, then the strand information in each window corresponds to the sum

of coverage coming from positive/negative reads; and not the number of posi-

tive/negative reads as default.

Value

A list of 2 numeric-Rle objects containing keeping probability of each +/- alignments. If getWin=TRUE then the list contains an additional DataFrame for the number of reads and coverage of the input window +/- alignments

.keptReadFragment

Calculate the read fragments to be kept

Description

Calculate the keeping probability of each read fragment based on the keeping probability of the windows containing it. Then get the list of read fragments to be kept.

Usage

.keptReadFragment(fragments, keptProbaW)

Arguments

fragments an IRanges object defining the starting, ending position of each fragment

keptProbaW an Rle object define the kept probability of each sliding window

Value

an integer vector of read fragment indices to be kept

.sequenceInfoInPartition

Calculate the first/last base/read of each sequence within each part of the partition.

Description

Calculate the first/last base/read of each sequence within each part of the partition.

Usage

.sequenceInfoInPartition(seqInfo, winWidth, winStep)

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Arguments

seqInfo a data frame that contains some key information of the sequences

winWidth the length of sliding window

winStep the step length to sliding the window

Value

Reduced sequence information (data.frame)

 $. \, {\tt summarize \ the \ histogram \ of \ strand \ proportions \ from \ the \ input \ win-}$

dows data frame

Description

Summarize the histogram of positive proportions from the input windows obtained from the function getStrandFromBamFile

Usage

```
.summarizeHist(
  windows,
  split = c(10L, 100L, 1000L),
  breaks = 100L,
  useCoverage = FALSE,
  groupBy = NULL,
  normalizeBy = NULL
)
```

Arguments

windows data frame containing the strand information of the sliding windows. Windows

can be obtained using the function getStrandFromBamFile.

split an integer vector that specifies how you want to partition the windows based

on the coverage. By default split = c(10,100,1000), which means that your windows will be partitionned into 4 groups, those have coverage < 10, from 10

to 100, from 100 to 1000, and > 1000

breaks an integer giving the number of bins for the histogram

useCoverage if TRUE then plot the coverage strand information, otherwise plot the number

of reads strand information. FALSE by default

groupBy the column names of windows that will be used to group the data

normalizeBy instead of using the raw read count/coverage, we will normalize it to a proportion

by dividing it to the total number of read count/coverage of windows that have

the same value in the normalizeBy columns.

Value

a dataframe object

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See Also

getStrandFromBamFile, plotHist, plotWin

checkPairedEnd

Test whether a bam file is single-end or paired-end

Description

Check the first 100000 first reads of the bam file to see whether it is single-end or paired-end

Usage

```
checkPairedEnd(file, yieldSize = 1e+05)
```

Arguments

file the input bam file. Your bamfile should be sorted and have an index file located

at the same path as well.

yieldSize the number of reads to be checked, 100000 by default.

Value

return TRUE if the input file is paired end, and FALSE if it is single end

Examples

```
file <- system.file('extdata','s1.sorted.bam',package = 'strandCheckR')
checkPairedEnd(file)</pre>
```

filterDNA

Filter reads comming from double strand sequences from a bam File

Description

Filter putative double strand DNA from a strand specific RNA-seq using a window sliding across the genome.

Usage

```
filterDNA(
  file,
  destination,
  statFile = "out.stat",
  sequences,
  mapqFilter = 0,
  paired,
  yieldSize = 1e+06,
  winWidth = 1000L,
  winStep = 100L,
```

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```
readProp = 0.5,
threshold = 0.7,
pvalueThreshold = 0.05,
useCoverage = FALSE,
mustKeepRanges,
getWin = FALSE,
minCov = 0,
maxCov = 0,
errorRate = 0.01
)
```

Arguments

file the input bam file to be filterd. Your bamfile should be sorted and have an index

file located at the same path.

destination the file path where the filtered output will be written

statFile the file to write the summary of the results

sequences the list of sequences to be filtered

mapqFilter every read that has mapping quality below mapqFilter will be removed before

any analysis. If missing, the entire bam file will be read.

paired if TRUE then the input bamfile will be considered as paired-end reads. If miss-

ing, 100 thousands first reads will be inspected to test if the input bam file in

paired-end or single-end.

yieldSize by default is 1e6, i.e. the bam file is read by block of reads whose size is defined

by this parameter. It is used to pass to same parameter of the scanBam function.

winWidth the length of the sliding window, 1000 by default.

winStep the step length to sliding the window, 100 by default.

readProp a read is considered to be included in a window if at least readProp of it is in

the window. Specified as a proportion. 0.5 by default.

threshold the strand proportion threshold to test whether to keep a window or not. 0.7 by

default

pvalueThreshold

the threshold for the p-value in the test of keeping windows. 0.05 by default

useCoverage if TRUE, then the strand information in each window corresponds to the sum

of coverage coming from positive/negative reads; and not the number of posi-

tive/negative reads as default.

mustKeepRanges a GRanges object; all reads that map to those ranges will be kept regardless the

strand proportion of the windows containing them.

getWin if TRUE, the function will not only filter the bam file but also return a data frame

containing the information of all windows of the original and filtered bam file.

minCov if useCoverage=FALSE, every window that has less than minCov reads will be

rejected regardless the strand proportion. If useCoverage=TRUE, every window

has max coverage least than minCov will be rejected. 0 by default

maxCov if useCoverage=FALSE, every window that has more than maxCov reads will

be kept regardless the strand proportion. If useCoverage=TRUE, every window with max coverage more than maxCov will be kept. If 0 then it doesn't have

effect on selecting window. 0 by default.

errorRate the probability that an RNA read takes the false strand. 0.01 by default.

Details

filterDNA reads a bam file containing strand specific RNA reads, and filter reads coming from putative double strand DNA. Using a window sliding across the genome, we calculate the positive/negative proportion of reads in each window. We then use logistic regression to estimate the strand proportion of reads in each window, and calculate the p-value when comparing that to a given threshold. Let π be the strand proportion of reads in a window.

Null hypothesis for positive window: $\pi \leq threshold$.

Null hypothesis for negative window: $\pi \geq 1 - threshold$.

Only windows with p-value <= pvalueThreshold are kept. For a kept positive window, each positive read in this window is kept with the probability (P-M)/P where P be the number of positive reads, and M be the number of negative reads. That is because those M negative reads are supposed to come from double-strand DNA, then there should be also M postive reads among the P positive reads come from double-strand DNA. In other words, there are only (P-M) positive reads come from RNA. Each negative read is kept with the probability equalling the rate that an RNA read of your sample has wrong strand, which is errorRate. Similar for kept negative windows.

Since each alignment can be belonged to several windows, then the probability of keeping an alignment is the maximum probability defined by all windows that contain it.

Value

if getWin is TRUE: a DataFrame object which could also be obtained by the function getStrandFromBamFile

See Also

```
getStrandFromBamFile, plotHist, plotWin
```

Examples

```
file <- system.file('extdata','s2.sorted.bam',package = 'strandCheckR')
out_bam <- tempfile(fileext = ".bam")
out_log <- tempfile(fileext = ".log")
filterDNA(file, sequences = '10', destination = out_bam, statFile = out_log)</pre>
```

getStrandFromBamFile Get the strand information of all windows from bam files

Description

Get the number of positive/negative reads/coverage of all slding windows from the bam input files

Usage

```
getStrandFromBamFile(
  files,
  sequences,
  mapqFilter = 0,
  yieldSize = 1e+06,
  winWidth = 1000L,
  winStep = 100L,
```

```
readProp = 0.5,
paired
)
```

Arguments

files the input bam files. Your bamfiles should be sorted and have their index files

located at the same path.

sequences character vector used to restrict analysed alignments to a subset of chromosomes

(i.e. sequences) within the provided bam file. These correspond to chromosomes/scaffolds of the reference genome to which the reads were mapped. If absent, the whole bam file will be read. NB: This must match the chromosomes as defined in your reference genome. If the reference chromosomes were specified using the 'chr' prefix, ensure the supplied vector matches this specification.

mapqFilter every read that has mapping quality below mapqFilter will be removed before

any analysis.

yieldSize by default is 1e6, i.e. the bam file is read by block of reads whose size is defined

by this parameter. It is used to pass to same parameter of the scanBam function.

winWidth the width of the sliding window, 1000 by default.
winStep the step length to sliding the window, 100 by default.

readProp A read is considered to be included in a window if at least readProp of it is in

the window. Specified as a proportion. 0.5 by default.

paired if TRUE then the input bamfile will be considered as paired-end reads. If miss-

ing, 100 thousands first reads will be inspected to test if the input bam file in

paired-end or single-end.

Details

This function moves along the specified chromosomes (i.e. sequences) using a sliding window approach, and counts the number of reads in each window which align to the +/- strands of the reference genome. As well as the number of reads, the total coverage for each strand is also returned for each window, representing the total number of bases covered.

Average coverage for the entire window can be simply calculated by dividing the total coverage by the window size.

Value

a DataFrame object containing the number of positive/negative reads and coverage of each window sliding across the bam file. The returned DataFrame has 10 columns:

Type: can be either SE if the input file contains single-end reads, or R1/R2 if the input file contains paired-end reads.

Seq: the reference sequence (chromosome/scaffold) that the reads were mapped to.

Start: the start position of the sliding window.

End: the end position of the sliding window.

NbPos/NbNeg: number of positive/negative reads that overlap the sliding window.

CovPos/CovNeg: number of bases coming from positive/negative reads that were mapped in the sliding window.

MaxCoverage: the maximum coverage within the sliding window.

File: the name of the input file.

See Also

```
filterDNA, plotHist, plotWin
```

Examples

```
file <- system.file('extdata','s1.sorted.bam',package = 'strandCheckR')
win <- getStrandFromBamFile(file,sequences='10')
win</pre>
```

getStrandFromReadInfo Get the strand information of all windows from read information

Description

Get the number of positive/negative reads of all windows from read information obtained from [Rsamtools::scanBam()]

Usage

```
getStrandFromReadInfo(
  readInfo,
  winWidth = 1000L,
  winStep = 100L,
  readProp = 0.5,
  subset = NULL
)
```

Arguments

```
readInfo a list containing read information returned by [Rsamtools::scanBam()].

winWidth the length of the sliding window, 1000 by default.

winStep the step length to sliding the window, 100 by default.

readProp A read is considered to be included in a window if at least readProp of it is in the window. Specified as a proportion. 0.5 by default.

subset an integer vector specifying the subset of reads to consider
```

Value

a DataFrame object containing the number of positive/negative reads and coverage of each window sliding .

See Also

```
filterDNA, getStrandFromBamFile
```

```
library(Rsamtools)
file <- system.file('extdata','s2.sorted.bam',package = 'strandCheckR')
readInfo <- scanBam(file, param =
ScanBamParam(what = c("pos","cigar","strand")))
getStrandFromReadInfo(readInfo[[1]],1000,100,0.5)</pre>
```

```
getWinOverlapEachIRange
```

Get the ranges of sliding windows that overlap each range of an IRanges object.

Description

Get the ranges of sliding windows that overlap each range of an IRanges object.

Usage

```
getWinOverlapEachIRange(
    x,
    winWidth = 1000L,
    winStep = 100L,
    readProp = 0.5,
    maxWin = Inf
)
```

Arguments

x	an IRanges object containing the start and end position of each read fragment.
winWidth	the width of the sliding window, 1000 by default.
winStep	the step length to sliding the window, 100 by default.
readProp	A read is considered to be included in a window if at least readProp of it is in the window. Specified as a proportion.
maxWin	The maximum window ID

Details

This finds the windows that overlap each range of the input IRanges object. Each range corresponds to a read fragment. This allows the total number of read fragments within a window to be calculated simply using [IRanges::coverage()].

Value

An IRanges object containing the index of the windows overlapping each read fragment

```
library(IRanges)
x <- IRanges(start=round(runif(100,1000,10000)),width=100)
getWinOverlapEachIRange(x)</pre>
```

```
getWinOverlapEachReadFragment
```

Get the window ranges that overlap each read fragment

Description

Calculate the window ranges that overlap each read fragment

Usage

```
getWinOverlapEachReadFragment(
  readInfo,
  strand,
  winWidth,
  winStep,
  readProp,
  useCoverage = FALSE,
  subset = NULL
)
```

Arguments

readInfo a list contains the read information

strand the considering strand

winWidth the width of the sliding window, 1000 by default.

winStep the step length to sliding the window, 100 by default.

readProp a read fragment is considered to be included in a window if and only if at least readProp percent of it is in the window.

useCoverage either base on coverage or number of reads

subset if we consider only a subset of the input reads

Value

If useCoverage=FALSE: an IRanges object which contains the range of sliding windows that overlap each read fragment. If useCoverage=TRUE: a list of two objects, the first one is the later IRanges object, the second one is an integer-Rle object which contains the coverage of the input readInfo

```
library(Rsamtools)
file <- system.file('extdata','s2.sorted.bam',package = 'strandCheckR')
readInfo <- scanBam(file, param =
ScanBamParam(what = c("pos","cigar","strand")))
getWinOverlapEachReadFragment(readInfo[[1]],"+",1000,100,0.5)</pre>
```

getWinOverlapGRanges Get the sliding windows that overlap a GRanges object

Description

Get the sliding windows that overlap a GRanges object.

Usage

```
getWinOverlapGRanges(
    x,
    seqInfo,
    winWidth = 1000L,
    winStep = 100L,
    nbOverlapBases = 1
)
```

Arguments

x a GRanges object, which defines the coordinates of the ranges in the reference

genome that all reads mapped to those ranges must be kept by the filtering

method filterDNA.

seqInfo a data frame that contains some key information of the sequences

winWidth the length of the sliding window, 1000 by default.
winStep the step length to sliding the window, 100 by default.

nbOverlapBases a window is considered to overlap with a range of x if it overlaps with at least

nbOverlapBases bases.

Details

This finds the windows that overlaps the positive/negative strand of a GRanges object. The GRanges object, which is mustKeepRanges in the filterDNA method, defines the coordinates of the ranges in the reference genome that all reads mapped to those ranges must be kept by the filtering method filterDNA. This method makes use of the method getWinOverlapEachIRange by pretending each given range as the range of a read. Since the widths of x are not necessarily the same (as normal read lengths), we use nbOverlapBases to specify the minimum number of bases that a window should overlap with a range of x, instead of using proprotion as readProp in getWinOverlapEachIRange.

Value

A list of two logical vectors (for positive and negative strand) defining which windows that overlap with the given GRanges object.

```
library(GenomicRanges)  
x \leftarrow GRanges(seqnames = "10", ranges = IRanges(start = c(10000,15000), end=c(20000,30000)), strand = c("+","-"))  
seqInfo \leftarrow data.frame("Sequence"=10,"FirstBaseInPart"=1)  
getWinOverlapGRanges(x,seqInfo)  
seqInfo \leftarrow data.frame("Sequence"=10,"FirstBaseInPart"=10000000)
```

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```
getWinOverlapGRanges(x,seqInfo)
```

Description

Intersect the windows with an annotation data frame to get features that overlap with each window

Usage

```
intersectWithFeature(
  windows,
  annotation,
  getFeatureInfo = FALSE,
  overlapCol = "OverlapFeature",
  mcolsAnnot,
  collapse,
  ...
)
```

Arguments

windows	data frame containing the strand information of the sliding windows. Windows can be obtained using the function getStrandFromBamFile.
annotation	a Grange object that you want to intersect with your windows. It can have mools which contains the information or features that could be able to integrate to the input windows
getFeatureInfo	whether to get the information of features in the mcols of annotation data or not. If FALSE the return windows will have an additional column indicating whether a window overlaps with any range of the annotion data. If TRUE the return windows will contain the information of features that overlap each window
overlapCol	the column name of the return windows indicating whether a window overlaps with any range of the annotion data.
mcolsAnnot	the column names of the mools of the annotation data that you want to get information
collapse	character which is used collapse multiple features that overlap with a same window into a string. If missing then we don't collapse them.
	used to pass parameters to GenomicRanges::findOverlaps

Value

the input windows DataFrame with some additional columns

See Also

```
{\tt getStrandFromBamFile}, {\tt plotHist}, {\tt plotWin}
```

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Examples

```
bamfilein = system.file('extdata','s2.sorted.bam',package = 'strandCheckR')
windows <- getStrandFromBamFile(file = bamfilein)
#add chr before chromosome names to be consistent with the annotation
windows$Seq <- paste0('chr',windows$Seq)
library(TxDb.Hsapiens.UCSC.hg38.knownGene)
annot <- transcripts(TxDb.Hsapiens.UCSC.hg38.knownGene)
# get the transcript names that overlap with each window
windows <- intersectWithFeature(windows,annot,mcolsAnnot='tx_name')
# just want to know whether there's any transcript that
# overlaps with each window
windows <- intersectWithFeature(windows,annot,overlapCol='OverlapTranscript')
plotHist(windows,facets = 'OverlapTranscript')
plotWin(windows,facets = 'OverlapTranscript')</pre>
```

plotHist

Plot the histogram of positive proportions

Description

Plot the histogram of positive proportions of the input data frame coming from getStrandFromBamFile

Usage

```
plotHist(
  windows,
  save = FALSE,
  file = "hist.pdf",
  groupBy = NULL,
  normalizeBy = NULL,
  split = c(10, 100, 1000),
  breaks = 100,
  useCoverage = FALSE,
  heatmap = FALSE,
  ...
)
```

Arguments

windows data frame containing the strand information of the sliding windows. Windows

can be obtained using the function getStrandFromBamFile.

save if TRUE, then the plot will be save into the file given by file parameter

file the file name to save to plot

groupBy the columns that will be used to split the data.

normalizeBy instead of using the raw read count/coverage, we will normalize it to a proportion

by dividing it to the total number of read count/coverage of windows that have

the same value in the normalizeBy columns.

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split an integer vector that specifies how you want to partition the windows based

on the coverage. By default split = c(10,100,1000), which means that your windows will be partitionned into 4 groups, those have coverage < 10, from 10

to 100, from 100 to 1000, and > 1000

breaks an integer giving the number of bins for the histogram

useCoverage if TRUE then plot the coverage strand information, otherwise plot the number

of reads strand information. FALSE by default

heatmap if TRUE, then use heat map to plot the histogram, otherwise use barplot. FALSE

by default.

... used to pass parameters to facet_wrap

Value

If heatmap=FALSE: a ggplot object

See Also

```
getStrandFromBamFile, plotWin
```

Examples

```
bamfilein = system.file('extdata','s1.sorted.bam',package = 'strandCheckR')
win <- getStrandFromBamFile(file = bamfilein,sequences='10')
plotHist(win)</pre>
```

plotWin

Plot the number of reads vs the proportion of '+' stranded reads.

Description

Plot the number of reads vs the proportion of '+' stranded reads of all windows from the input data frame.

Usage

```
plotWin(
    windows,
    split = c(10, 100, 1000),
    threshold = c(0.6, 0.7, 0.8, 0.9),
    save = FALSE,
    file = "win.pdf",
    groupBy = NULL,
    useCoverage = FALSE,
    ...
)
```

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Arguments

windows	data frame containing the strand information of the sliding windows. Windows should be obtained using the function <code>getStrandFromBamFile</code> to ensure the correct data structure.
split	an integer vector that specifies how you want to partition the windows based on coverage. By default $split = c(10,100,1000)$, partition windows into 4 groups based on these values.
threshold	a numeric vector between 0.5 & 1 that specifies which threshold lines to draw on the plot. The positive windows above the threshold line (or negative windows below the threshold line) will be kept when using filterDNA.
save	if TRUE, then the plot will be save into the file given by file parameter
file	the file name to save to plot
groupBy	the column that will be used to split the data (which will be used in the facets method of ggplot2).
useCoverage	if TRUE then plot the coverage strand information, otherwise plot the number of reads strand information. FALSE by default
	used to pass parameters to facet_wrap during plotting

Details

This function will plot the proportion of '+' stranded reads for each window, against the number of reads in each window. The threshold lines indicate the hypothetical boundary where windows will contain reads to kept or discarded using the filtering methods of filterDNA. Any plot can be easily modified using standard ggplot2 syntax (see Examples)

Value

The plot will be returned as a standard ggplot2 object

See Also

```
{\tt getStrandFromBamFile}, {\tt plotHist}
```

```
bamfilein = system.file('extdata','s2.sorted.bam',package = 'strandCheckR')
windows <- getStrandFromBamFile(file = bamfilein,sequences = '10')
plotWin(windows)

# Change point colour using ggplot2
library(ggplot2)
plotWin(windows) +
scale_colour_manual(values = rgb(seq(0, 1, length.out = 4), 0, 0))</pre>
```

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