

# Package ‘inti’

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**Type** Package

**Version** 0.6.8

**Title** Tools and Statistical Procedures in Plant Science

**Description** The 'inti' package is part of the 'inkaverse' project for developing different procedures and tools used in plant science and experimental designs. The main aim of the package is to support researchers during the planning of experiments and data collection (`tarpu()`), data analysis and graphics (`yupana()`), and scientific writing.

Learn more about the 'inkaverse' project at [<https://inkaverse.com/>](https://inkaverse.com/).

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**License** GPL-3 | file LICENSE

**URL** <https://inkaverse.com/>, <https://github.com/flavjack/inti>

**BugReports** <https://github.com/flavjack/inti/issues/>

**Depends** shiny, ggplot2, dplyr, tidyr, tibble, R (>= 4.1.0)

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---

`colortext`*Colourise text for display in the terminal*

---

**Description**

If R is not currently running in a system that supports terminal colours the text will be returned unchanged.

**Usage**

```
colortext(text, fg = "red", bg = NULL)
```

**Arguments**

<code>text</code>	character vector
<code>fg</code>	foreground colour, defaults to white
<code>bg</code>	background colour, defaults to transparent

**Details**

Allowed colours are: black, blue, brown, cyan, dark gray, green, light blue, light cyan, light gray, light green, light purple, light red, purple, red, white, yellow

**Author(s)**

testthat package

**Examples**

```
print(colortext("Red", "red"))
cat(colortext("Red", "red"), "\n")
cat(colortext("White on red", "white", "red"), "\n")
```

---

`design_noreps`*Experimental design without replications*

---

**Description**

Function to deploy field-book experiment without replications

**Usage**

```
design_noreps(
  factors,
  type = "sorted",
  zigzag = FALSE,
  nrows = NA,
  serie = 100,
  seed = NULL,
  ffname = "inkaverse",
  qrcode = "{ffname}{plots}{factors}"
)
```

**Arguments**

factors	Lists with names and factor vector [list].
type	Randomization in the list [string: sorted, unsorted]
zigzag	Experiment layout in zigzag [logic: FALSE].
nrows	Experimental design dimension by rows [numeric: value]
serie	Number to start the plot id [numeric: 1000].
seed	Replicability from randomization [numeric: NULL].
ffname	Bar code prefix for data collection [string: "inkaverse"].
qrcode	Concatenate the QR code [string: {ffname}{plots}{factors}]

**Value**

A list with the field-book design and parameters

**Examples**

```
## Not run:

library(inti)

factores <- list("geno" = c(1:99))

fb <- design_noreps(factors = factores
  , type = "sorted"
  , zigzag = F
  , nrows = 10
  )

dsg <- fb$fieldbook

fb %>%
  tarpuy_plotdesign(fill = "plots")

fb$parameters
```

```
## End(Not run)
```

---

design_repblock	<i>Experimental design in CRD and RCBD</i>
-----------------	--

---

## Description

Function to deploy field-book experiment for CRD and RCBD

## Usage

```
design_repblock(
  nfactors = 1,
  factors,
  type = "crd",
  rep = 3,
  zigzag = FALSE,
  nrows = NA,
  serie = 100,
  seed = NULL,
  fbname = "inkaverse",
  qrcode = "{fbname}{plots}{factors}"
)
```

## Arguments

nfactors	Number of factor in the experiment [numeric: 1].
factors	Lists with names and factor vector [list].
type	Type of experimental arrange [string: "crd" "rcbd" "lsd"]
rep	Number of replications in the experiment [numeric: 3].
zigzag	Experiment layout in zigzag [logic: F].
nrows	Experimental design dimension by rows [numeric: value]
serie	Number to start the plot id [numeric: 100].
seed	Replicability from randomization [numeric: NULL].
fbname	Bar code prefix for data collection [string: "inkaverse"].
qrcode	Concatenate the QR code [string: {fbname}{plots}{factors}]

## Value

A list with the field-book design and parameters

**Examples**

```

## Not run:

library(inti)

factores <- list("geno" = c("A", "B", "C", "D", "D", 1, NA, NA, NULL, "NA")
, "salt stress" = c(0, 50, 200, 200, "T0", NA, NULL, "NULL")
, time = c(30, 60, 90)
)

fb <- design_repblock(nfactors = 2
, factors = factores
, type = "rcbd"
, rep = 5
, zigzag = T
, seed = 0
, nrows = 20
, qrcode = "{fbname}{plots}{factors}"
)

dsg <- fb$fieldbook

fb %>%
  tarpuy_plotdesign(fill = "plots")

fb$parameters

## End(Not run)

```

---

figure2qmd

*Figure to Quarto format*


---

**Description**

Use Articul8 Add-ons from Google docs to build Ricles

**Usage**

```
figure2qmd(text, path = ".", opts = NA)
```

**Arguments**

text	Markdown text with figure information [string]
path	Image path for figures [path: "." (base directory)]
opts	chunk options in brackets [string: NA]

**Details**

Quarto option can be included in the title using "{{ }}" separated by commas

**Value**

string mutated

---

figure2rmd	<i>Figure to Rmarkdown format</i>
------------	-----------------------------------

---

**Description**

Use Articul8 Add-ons from Google docs to build Rarticles

**Usage**

```
figure2rmd(text, path = ".", opts = NA)
```

**Arguments**

text	String with the table information
path	Path of the image for the figure
opts	chunk options in brackets.

**Value**

Mutated string

---

footnotes	<i>Footnotes in tables</i>
-----------	----------------------------

---

**Description**

Include tables footnotes and symbols for kables in pandoc format

**Usage**

```
footnotes(table, notes = NULL, label = "Note:", notation = "alphabet")
```

**Arguments**

table	Kable output in pandoc format.
notes	Footnotes for the table.
label	Label for start the footnote.
notation	Notation for the footnotes (default = "alphabet"). See details.

**Details**

You should use the pandoc format `kable(format = "pipe")`. You can add the footnote symbol using `{hyphen}` in your table. notation could be use: "alphabet", "number", "symbol", "none".

**Value**

Table with footnotes for word and html documents

---

gdoc2qmd

*Google docs to Rmarkdown*


---

**Description**

Use Articul8 Add-ons from Google docs to build Ricles

**Usage**

```
gdoc2qmd(file, export = NA, format = "qmd", type = "asis", fill_table = "down")
```

**Arguments**

<code>file</code>	Zip file path from Articul8 exported in md format [path]
<code>export</code>	Path to export the files [path: NA (file directory)]
<code>format</code>	Output format [string: "qmd" "rmd"]
<code>type</code>	output file type [strig: "asis" "list", "listfull", "full"]
<code>fill_table</code>	extract table in listfull [strig: "up", "down"]

**Details**

Document rendering until certain point: "#! end" Include for next page: "#! newpage" You can include the cover page params using "#!" in a Google docs table

**Value**

path



---

H2cal

*Broad-sense heritability in plant breeding*


---

### Description

Heritability in plant breeding on a genotype difference basis

### Usage

```
H2cal(
  data,
  trait,
  gen.name,
  rep.n,
  env.n = 1,
  year.n = 1,
  env.name = NULL,
  year.name = NULL,
  fixed.model,
  random.model,
  summary = FALSE,
  emmeans = FALSE,
  weights = NULL,
  plot_diag = FALSE,
  outliers.rm = FALSE,
  trial = NULL
)
```

### Arguments

<code>data</code>	Experimental design data frame with the factors and traits.
<code>trait</code>	Name of the trait.
<code>gen.name</code>	Name of the genotypes.
<code>rep.n</code>	Number of replications in the experiment.
<code>env.n</code>	Number of environments (default = 1). See details.
<code>year.n</code>	Number of years (default = 1). See details.
<code>env.name</code>	Name of the environments (default = NULL). See details.
<code>year.name</code>	Name of the years (default = NULL). See details.
<code>fixed.model</code>	The fixed effects in the model (BLUEs). See examples.
<code>random.model</code>	The random effects in the model (BLUPs). See examples.
<code>summary</code>	Print summary from random model (default = FALSE).
<code>emmeans</code>	Use emmeans for calculate the BLUEs (default = FALSE).
<code>weights</code>	an optional vector of 'prior weights' to be used in the fitting process (default = NULL).

<code>plot_diag</code>	Show diagnostic plots for fixed and random effects (default = FALSE). Options: "base", "ggplot". .
<code>outliers.rm</code>	Remove outliers (default = FALSE). See references.
<code>trial</code>	Column with the name of the trial in the results (default = NULL).

### Details

The function allows to made the calculation for individual or multi-environmental trials (MET) using fixed and random model.

1. The variance components based in the random model and the population summary information based in the fixed model (BLUEs).
2. Heritability under three approaches: Standard (ANOVA), Cullis (BLUPs) and Piepho (BLUEs).
3. Best Linear Unbiased Estimators (BLUEs), fixed effect.
4. Best Linear Unbiased Predictors (BLUPs), random effect.
5. Table with the outliers removed for each model.

For individual experiments is necessary provide the `trait`, `gen.name`, `rep.n`.

For MET experiments you should `env.n` and `env.name` and/or `year.n` and `year.name` according your experiment.

The BLUEs calculation based in the pairwise comparison could be time consuming with the increase of the number of the genotypes. You can specify `emmeans = FALSE` and the calculate of the BLUEs will be faster.

If `emmeans = FALSE` you should change 1 by 0 in the fixed model for exclude the intersect in the analysis and get all the genotypes BLUEs.

For more information review the references.

### Value

list

### Author(s)

Maria Belen Kistner

Flavio Lozano Isla

### References

Bernal Vasquez, Angela Maria, et al. "Outlier Detection Methods for Generalized Lattices: A Case Study on the Transition from ANOVA to REML." *Theoretical and Applied Genetics*, vol. 129, no. 4, Apr. 2016.

Buntaran, H., Piepho, H., Schmidt, P., Ryden, J., Halling, M., and Forkman, J. (2020). Cross validation of stagewise mixed model analysis of Swedish variety trials with winter wheat and spring barley. *Crop Science*, 60(5).

Schmidt, P., J. Hartung, J. Bennewitz, and H.P. Piepho. 2019. Heritability in Plant Breeding on a Genotype Difference Basis. *Genetics* 212(4).

Schmidt, P., J. Hartung, J. Rath, and H.P. Piepho. 2019. Estimating Broad Sense Heritability with Unbalanced Data from Agricultural Cultivar Trials. *Crop Science* 59(2).

Tanaka, E., and Hui, F. K. C. (2019). Symbolic Formulae for Linear Mixed Models. In H. Nguyen (Ed.), *Statistics and Data Science*. Springer.

Zystro, J., Colley, M., and Dawson, J. (2018). Alternative Experimental Designs for Plant Breeding. In *Plant Breeding Reviews*. John Wiley and Sons, Ltd.

## Examples

```
library(inti)

dt <- potato

hr <- H2cal(data = dt
            , trait = "stemdw"
            , gen.name = "geno"
            , rep.n = 5
            , fixed.model = "0 + (1|bloque) + geno"
            , random.model = "1 + (1|bloque) + (1|geno)"
            , emmeans = TRUE
            , plot_diag = FALSE
            , outliers.rm = TRUE
            )

hr$tabsmr
hr$blues
hr$blups
hr$outliers
```

---

include\_pdf

*Include PDF in markdown documents*

---

## Description

Insert PDF files in markdown documents

## Usage

```
include_pdf(file, width = "100%", height = "600")
```

## Arguments

file	file path from pdf file.
width	width preview file.
height	height preview file.

**Value**

html code for markdown

---

include_table	<i>Table with footnotes</i>
---------------	-----------------------------

---

**Description**

Include tables with title and footnotes for word and html documents

**Usage**

```
include_table(table, caption = NA, notes = NA, label = NA, notation = "none")
```

**Arguments**

table	Data frame.
caption	Table caption (default = NULL). See details.
notes	Footnotes for the table (default = NA). See details.
label	Label for start the footnote (default = NA).
notation	Notation for the symbols and footnotes (default = "none") Others: "alphabet", "number", "symbol".

**Value**

Table with caption and footnotes

**Examples**

```
library(inti)

table <- data.frame(
  x = rep_len(1, 5)
  , y = rep_len(3, 5)
  , z = rep_len("c", 5)
)

table %>% inti::include_table(
  caption = "Title caption b) line 0
a) line 1
b) line 2"
  , notes = "Footnote"
  , label = "Where:"
)
```

---

`jc_tombola`*Journal Club Tombola*

---

**Description**

Function for arrange journal club schedule

**Usage**

```
jc_tombola(  
  data,  
  members,  
  papers = 1,  
  group = NA,  
  gr_lvl = NA,  
  status = NA,  
  st_lvl = "active",  
  frq = 7,  
  date = NA,  
  seed = NA  
)
```

**Arguments**

<code>data</code>	Data frame with the members and their information.
<code>members</code>	Columns with the members names.
<code>papers</code>	Number of paper by meeting
<code>group</code>	Column for arrange the group.
<code>gr_lvl</code>	Levels in the groups for the arrange. See details.
<code>status</code>	Column with the status of the members.
<code>st_lvl</code>	Level to confirm the assistance in the JC. See details.
<code>frq</code>	Number of the day for each session.
<code>date</code>	Date when start the first session of JC.
<code>seed</code>	Number for replicate the results (default = date).

**Details**

The function could consider n levels for `gr_lvl`. In the case of more levels using "both" or "all" will be the combination. The suggested levels for `st_lvl` are: active or spectator. Only the "active" members will enter in the schedule.

**Value**

data frame with the schedule for the JC

---

mean_comparison	<i>Mean comparison test</i>
-----------------	-----------------------------

---

### Description

Function to compare treatment from lm or aov using data frames

### Usage

```
mean_comparison(
  data,
  response,
  model_factors,
  comparison,
  test_comp = "SNK",
  sig_level = 0.05
)
```

### Arguments

data	Fieldbook data.
response	Model used for the experimental design.
model_factors	Factor in the model.
comparison	Significance level for the analysis (default = 0.05).
test_comp	Comparison test (default = "SNK"). Others: "TUKEY", "DUNCAN".
sig_level	Significance level for the analysis (default = 0.05).

### Value

list

### Examples

```
## Not run:

library(inti)
library(gsheets)

url <- paste0("https://docs.google.com/spreadsheets/d/"
             , "15r7ZwcZZHbEg1t1F6gSFvCTFA-CFzVBWwg3mFlRyKPs/"
             , "edit#gid=172957346")
# browseURL(url)

fb <- gsheets2tbl(url)

mc <- mean_comparison(data = fb
                     , response = "spad_29")
```

```

, model_factors = "bloque* geno*treat"
, comparison = c("geno", "treat")
, test_comp = "SNK"
)
mc$comparison
mc$stat

## End(Not run)

```

met

*Swedish cultivar trial data*

## Description

The datasets were obtained from official Swedish cultivar tests. Dry matter yield was analyzed. All trials were laid out as alpha-designs with two replicates. Within each replicate, there were five to seven incomplete blocks.

## Usage

```
met
```

## Format

A data frame with 1069 rows and 8 variables:

**zone** Sweden is divided into three different agricultural zones: South, Middle, and North

**location** Locations: 18 location in the Zones

**rep** Replications (4): number of replication in the experiment

**alpha** Incomplete blocks (8) in the alpha-designs

**cultivar** Cultivars (30): genotypes evaluated

**yield** Yield in kg/ha

**year** Year (1): 2016

**env** enviroment (18): combination zone + location + year

## Source

[doi:10.1002/csc2.20177](https://doi.org/10.1002/csc2.20177)

---

metamorphosis	<i>Transform fieldbooks based in a dictionary</i>
---------------	---

---

### Description

Transform entire fieldbook according to data a dictionary

### Usage

```
metamorphosis(fieldbook, dictionary, from, to, index, colnames)
```

### Arguments

fieldbook	Data frame with the original information.
dictionary	Data frame with new names and categories. See details.
from	Column of the dictionary with the original names.
to	Column of the dictionary with the new names.
index	Column of the dictionary with the type and level of the variables.
colnames	Character vector with the name of the columns.

### Details

The function require at least three columns.

1. Original names (from).
2. New names (to).
3. Variable type (index).

### Value

List with two objects. 1. New data frame. 2. Dictionary.

---

outliers_remove	<i>Remove outliers</i>
-----------------	------------------------

---

### Description

Use the method M4 in Bernal Vasquez (2016). Bonferroni Holm test to judge residuals standardized by the re scaled MAD (BH MADR).

### Usage

```
outliers_remove(data, trait, model, drop_na = TRUE)
```



**Arguments**

data	Experimental design data frame with the factors and traits.
trait	Name of the trait.
model	The fixed or random effects in the model.
drop_na	drop NA values from the data.frame

**Details**

Function to remove outliers in MET experiments

**Value**

list. 1. Table with data without outliers. 2. The outliers in the dataset.

**References**

Bernal Vasquez, Angela Maria, et al. "Outlier Detection Methods for Generalized Lattices: A Case Study on the Transition from ANOVA to REML." *Theoretical and Applied Genetics*, vol. 129, no. 4, Apr. 2016.

**Examples**

```
library(inti)

rmout <- potato %>% outliers_remove(
  data = .
  , trait = "stemdw"
  , model = "0 + treat*geno + (1|bloque)"
  , drop_na = FALSE
)

rmout
```

---

plot\_diag

*Diagnostic plots*

---

**Description**

Function to plot the diagnostic of models

**Usage**

```
plot_diag(model, title = NA)
```

**Arguments**

model	Statistical model
title	Plot title

**Value**

plots

**Examples**

```
## Not run:

library(inti)

lm <- aov(stemdw ~ bloque + geno*treat, data = potato)

# lm <- potato %>% lme4::lmer(stemdw ~ (1|bloque) + geno*treat, data = .)

plot(lm, which = 1)
plot_diag(lm)[3]

plot(lm, which = 2)
plot_diag(lm)[2]

plot(lm, which = 3)
plot_diag(lm)[4]

plot(lm, which = 4)
plot_diag(lm)[1]

## End(Not run)
```

---

plot_diagnostic	<i>Diagnostic plots</i>
-----------------	-------------------------

---

**Description**

Function to plot the diagnostic of models

**Usage**

```
plot_diagnostic(data, formula, title = NA)
```

**Arguments**

data	Experimental design data frame with the factors and traits.
formula	Mixed model formula
title	Plot title

**Value**

plots

**Examples**

```
## Not run:  
  
library(inti)  
  
plot_diagnostic(data = potato  
                , formula = stemdw ~ (1|bloque) + geno*treat)  
  
## End(Not run)
```

---

plot\_raw

*Plot raw data*

---

**Description**

Function use the raw data for made a boxplot graphic

**Usage**

```
plot_raw(  
  data,  
  type = "boxplot",  
  x,  
  y,  
  group = NULL,  
  xlab = NULL,  
  ylab = NULL,  
  glab = NULL,  
  ylimits = NULL,  
  xlimits = NULL,  
  xrotation = NULL,  
  legend = "top",  
  xtext = NULL,  
  gtext = NULL,  
  color = TRUE,  
  linetype = 1,  
  opt = NULL  
)
```

**Arguments**

data	raw data
type	Type of graphic. "boxplot" or "scatterplot"
x	Axis x variable
y	Axis y variable
group	Group variable
xlab	Title for the axis x
ylab	Title for the axis y
glab	Title for the legend
ylimits	Limits and break of the y axis c(initial, end, brakes)
xlimits	For scatter plot. Limits and break of the x axis c(initial, end, brakes)
xrotation	Rotation in x axis c(angle, h, v)
legend	the position of legends ("none", "left", "right", "bottom", "top", or two-element numeric vector)
xtext	Text labels in x axis using a vector
gtext	Text labels in groups using a vector
color	Colored figure (TRUE), black & white (FALSE) or color vector
linetype	Line type for regression. Default = 0
opt	Add new layers to the plot

**Details**

You could add additional layer to the plot using "+" with ggplot2 options

**Value**

plot

**Examples**

```
## Not run:

library(inti)

fb <- potato

fb %>%
  plot_raw(type = "box"
            , x = "geno"
            , y = "twue"
            , group = NULL
            , ylab = NULL
            , xlab = NULL
            , glab = ""
            )
```

```
fb %>%
  plot_raw(type = "sca"
           , x = "geno"
           , y = "twue"
           , group = "treat"
           , color = c("red", "blue")
           )

## End(Not run)
```

---

plot\_smr

*Plot summary data*

---

### Description

Graph summary data into bar o line plot

### Usage

```
plot_smr(
  data,
  type = NULL,
  x = NULL,
  y = NULL,
  group = NULL,
  xlab = NULL,
  ylab = NULL,
  glab = NULL,
  ylimits = NULL,
  xrotation = c(0, 0.5, 0.5),
  xtext = NULL,
  gtext = NULL,
  legend = "top",
  sig = NULL,
  sigsize = 3,
  error = NULL,
  color = TRUE,
  opt = NULL
)
```

### Arguments

data	Output from summary data
type	Type of graphic. "bar" or "line"
x	Axis x variable

y	Axis y variable
group	Group variable
xlab	Title for the axis x
ylab	Title for the axis y
glab	Title for the legend
ylimits	limits of the y axis c(initial, end, brakes)
xrotation	Rotation in x axis c(angle, h, v)
xtext	Text labels in x axis using a vector
gtext	Text labels in group using a vector
legend	the position of legends ("none", "left", "right", "bottom", "top", or two-element numeric vector)
sig	Column with the significance
sigsize	Font size in significance letters
error	Show the error bar ("ste" or "std")
color	colored figure (TRUE), black & white (FALSE) or color vector
opt	Add news layer to the plot

### Details

If the table is a out put of mean\_comparison(graph\_opts = TRUE) function. Its contain all the parameter for the plot.

You could add additional layer to the plot using "+" with ggplot2 options

### Value

plot

### Examples

```
## Not run:

library(inti)

fb <- potato

yrs <- yupana_analysis(data = fb
                      , response = "lfa"
                      , model_factors = "geno*treat"
                      , comparison = c("geno", "treat")
                      )

yrs$meancomp %>%
  plot_smr(type = "bar"
          , x = "geno"
          , y = "lfa"
          , xlab = "")
```

```

, group = "treat"
, glab = "Tratamientos"
, error = "ste"
, sig = "sig"
#, ylimits = c(0, 1, 0.2)
, color = c("red", "black")
, gtext = c("Irrigado", "Sequia")
)

## End(Not run)

```

---

potato

*Water use efficiency in 15 potato genotypes*

---

### Description

Experiment to evaluate the physiological response from 15 potatoes genotypes under water deficit condition. The experiment had a randomized complete block design with five replications. The stress started at 30 day after planting.

### Usage

potato

### Format

A data frame with 150 rows and 17 variables:

**treat** Water deficit treatments: sequia, irrigado

**geno** 15 potato genotypes

**bloque** blocks for the experimentl design

**spad\_29** Relative chlorophyll content (SPAD) at 29 day after planting

**spad\_83** Relative chlorophyll content (SPAD) at 84 day after planting

**rwc\_84** Relative water content (percentage) at 84 day after planting

**op\_84** Osmotic potential (Mpa) at 84 day after planting

**leafdw** leaf dry weight (g)

**stemdw** stem dry weight (g)

**rootdw** root dry weight (g)

**tubdw** tuber dry weight (g)

**biomdw** total biomass dry weight (g)

**hi** harvest index

**ttrans** total transpiration (l)

**wue** water use efficiency (g/l)  
**twue** tuber water use efficiency (g/l)  
**lfa** leaf area (cm<sup>2</sup>)

---

remove\_outliers      *Remove outliers using mixed models*

---

### Description

Use the method M4 in Bernal Vasquez (2016). Bonferroni Holm test to judge residuals standardized by the re scaled MAD (BH MADR).

### Usage

```
remove_outliers(data, formula, drop_na = FALSE, plot_diag = FALSE)
```

### Arguments

data	Experimental design data frame with the factors and traits.
formula	mixed model formula.
drop_na	drop NA values from the data.frame
plot_diag	Diagnostic plot based in the raw and clean data

### Details

Function to remove outliers in MET experiments

### Value

list. 1. Table with date without outliers. 2. The outliers in the dataset.

### References

Bernal Vasquez, Angela Maria, et al. "Outlier Detection Methods for Generalized Lattices: A Case Study on the Transition from ANOVA to REML." *Theoretical and Applied Genetics*, vol. 129, no. 4, Apr. 2016.

### Examples

```
library(inti)

rmout <- potato %>%
  remove_outliers(data = .
    , formula = stemdw ~ 0 + (1|bloque) + treat*geno
    , plot_diag = FALSE
    , drop_na = FALSE
  )
```



rmout

---

split\_folder

*Split folder*

---

### Description

Function to split folder by size or number of elements

### Usage

```
split_folder(  
  folder,  
  export,  
  units = "megas",  
  size = 500,  
  zip = TRUE,  
  remove = FALSE  
)
```

### Arguments

folder	Path of folder to split (path).
export	Path to export the split folders (path).
units	Units to split folder (string: "megas", "number").
size	Folder size by the units selected (numeric).
zip	Zip split folders (logical).
remove	Remove the split folder after zip (logical).

### Value

zip files

### Examples

```
## Not run:  
  
split_folder("pictures/QUINOA 2018-2019 SC SEEDS EDWIN - CAMACANI/"  
  , "pictures/split_num", remove = T, size = 400, units = "number")  
  
## End(Not run)
```

---



*Table to Quarto format*


---

**Description**

Use Articul8 Add-ons from Google docs to build Rarticles

**Usage**

```
table2qmd(text, type = "asis")
```

**Arguments**

text	Markdown text with table information (string)
type	output file type [strig: "asis" "list", "listfull", "full"]

**Value**

string mutated

---

table2rmd

*Table to Rmarkdown format*


---

**Description**

Use Articul8 Add-ons from Google docs to build Rarticles

**Usage**

```
table2rmd(text, opts = NA)
```

**Arguments**

text	String with the table information
opts	chunk options in brackets.

**Value**

Mutated string

---

tarpuy	<i>Interactive fieldbook designs</i>
--------	--------------------------------------

---

**Description**

Invoke RStudio addin to create fieldbook designs

**Usage**

```
tarpuy(dependencies = FALSE)
```

**Arguments**

dependencies    Install package dependencies for run the app

**Details**

Tarpuy allow to create experimental designs under an interactive app.

**Value**

Shiny app

**Examples**

```
if(interactive()){  
  inti::tarpuy()  
}
```

---

tarpuy_design	<i>Fieldbook experimental designs</i>
---------------	---------------------------------------

---

**Description**

Function to deploy experimental designs

**Usage**

```
tarpuy_design(
  data,
  nfactors = 1,
  type = "crd",
  rep = 2,
  zigzag = FALSE,
  nrows = NA,
  serie = 100,
  seed = NULL,
  fbname = NA,
  qrcode = "{fbname}{plots}{factors}"
)
```

**Arguments**

data	Experimental design data frame with the factors and level. See examples.
nfactors	Number of factor in the experiment(default = 1). See details.
type	Type of experimental arrange (default = "crd"). See details.
rep	Number of replications in the experiment (default = 3).
zigzag	Experiment layout in zigzag [logic: FALSE].
nrows	Experimental design dimension by rows [numeric: value]
serie	Number to start the plot id [numeric: 100].
seed	Replicability of draw results (default = 0) always random. See details.
fbname	Barcode prefix for data collection.
qrcode	[string: {fbname}{plots}{factors}] String to concatenate the QR code.

**Details**

The function allows to include the arguments in the sheet that have the information of the design. You should include 2 columns in the sheet: {arguments} and {values}. See examples. The information will be extracted automatically and deploy the design. nfactors = 1: crd, rcbd, lsd, lattice. nfactors = 2 (factorial): split-crd, split-rcbd split-lsd nfactors >= 2 (factorial): crd, rcbd, lsd.

**Value**

A list with the fieldbook design

**Examples**

```
## Not run:

library(inti)
library(gsheets)
```

```
url <- paste0("https://docs.google.com/spreadsheets/d/"
             , "1510f0Kj0g4CDEAFkrpFbr-zNMn1e_Hou90_wuf7Vdo4/edit?gid=1479851579#gid=1479851579")
# browseURL(url)

fb <- gsheets2tbl(url)

dsg <- fb %>% tarpuy_design()

dsg %>%
  tarpuy_plotdesign()

## End(Not run)
```

---

tarpuy\_plex

*Fieldbook plan information*

---

## Description

Information for build a plan for an experiment (PLEX)

## Usage

```
tarpuy_plex(
  data = NULL,
  title = NULL,
  objectives = NULL,
  hypothesis = NULL,
  rationale = NULL,
  references = NULL,
  plan = NULL,
  institutions = NULL,
  researchers = NULL,
  manager = NULL,
  location = NULL,
  altitude = NULL,
  georeferencing = NULL,
  environment = NULL,
  start = NA,
  end = NA,
  about = NULL,
  fieldbook = NULL,
  project = NULL,
  repository = NULL,
  manuscript = NULL,
  album = NULL,
  nfactor = 2,
  design = "rcbd",
```

```

    rep = 3,
    zigzag = FALSE,
    nrows = NA,
    serie = 100,
    seed = 0,
    qrcode = "{fbname}{plots}{factors}"
  )

```

### Arguments

data	Data with the fieldbook information.
title	Project title.
objectives	The objectives of the project.
hypothesis	What are the expected results.
rationale	Based in which evidence is planned the experiment.
references	References.
plan	General description of the project (M & M).
institutions	Institutions involved in the project.
researchers	Persons involved in the project.
manager	Persons responsible of the collection of the data.
location	Location of the project.
altitude	Altitude of the experiment (m.a.s.l).
georeferencing	Georeferencing information.
environment	Environment of the experiment (greenhouse, lab, etc).
start	The date of the start of the experiments.
end	The date of the end of the experiments.
about	Short description of the project.
fieldbook	Name or ID for the fieldbook/project.
project	link for project.
repository	link to the repository.
manuscript	link for manuscript.
album	link with the photos of the project.
nfactor	Number of factors for the design.
design	Type of design.
rep	Number of replication.
zigzag	Experiment layout in zigzag [logic: F]
nrows	Experimental design dimension by rows [numeric: value]
serie	Number of digits in the plots.
seed	Seed for the randomization.
qrcode	Concatenate the QR code [string: {fbname}{plots}{factors}]

**Details**

Provide the information available.

**Value**

data frame or list of arguments:

1. info
2. variables
3. design
4. logbook
5. timetable
6. budget

---

tarpuv_plotdesign	<i>Fieldbook plot experimental designs</i>
-------------------	--

---

**Description**

Plot fieldbook sketch designs based in experimental design

**Usage**

```
tarpuv_plotdesign(
  data,
  factor = NA,
  fill = "plots",
  xlab = NULL,
  ylab = NULL,
  glab = NULL
)
```

**Arguments**

<code>data</code>	Experimental design data frame with the factors and level. See examples.
<code>factor</code>	Vector with the name of the columns with the factors.
<code>fill</code>	Value for fill the experimental units (default = "plots").
<code>xlab</code>	Title for x axis.
<code>ylab</code>	Title for y axis.
<code>glab</code>	Title for group axis.

**Details**

The function allows to plot the experimental design according the field experiment design.

**Value**

plot

**Examples**

```
## Not run:

library(inti)
library(gsheets)

url <- paste0("https://docs.google.com/spreadsheets/d/"
             , "1_BVzChX_-lzXhB7HAm6FeSrwq9iKfZ39_Sl8NFC6k7U/edit#gid=1834109539")
# browseURL(url)

fb <- gsheets2tbl(url)

dsg <- fb %>% tarpuy_design()

dsg

dsg %>% str()

dsg %>%
  tarpuy_plotdesign()

## End(Not run)
```

---

tarpuy\_traits

*Field book traits*


---

**Description**

Function to export field book and traits for be used in field book app.

**Usage**

```
tarpuy_traits(fieldbook = NULL, last_factor = NULL, traits = NULL)
```

**Arguments**

fieldbook	Experiment field book [dataframe].
last_factor	Last factor in the field book [string: colnames]
traits	Traits information [dataframe or list].

**Details**

For the traits parameters you can used shown in the Field Book app



**Value**

list

**Examples**

```
library(inti)

fieldbook <- inti::potato

traits <- list(
  list(variable = "altura de planta"
        , trait = "altp"
        , format = "numeric"
        , when = "30, 40, 50"
        , samples = 3
        , units = "cm"
        , details = NA
        , minimum = 0
        , maximum = 100
        )
  , list(variable = "severidad"
        , trait = "svr"
        , format = "scategorical"
        , when = "30, 40, 50"
        , samples = 1
        , units = "scale"
        , details = NA
        , categories = "1, 3, 5, 7, 9"
        )
  , list(variable = "foto"
        , trait = "foto"
        , format = "photo"
        , when = "hrv, pshrv"
        , samples = 1
        , units = "image"
        , details = NA
        )
  , list(variable = "germinacion"
        , trait = "ger"
        , format = "boolean"
        , when = "30, 40, 50"
        , samples = 1
        , units = "logical"
        , details = NA
        )
)

fbapp <- tarpuy_traits(fieldbook, last_factor = "bloque", traits)

## Not run:

library(inti)
```

```
library(gsheets)

url_fb <- paste0("https://docs.google.com/spreadsheets/d/"
  , "1510f0Kj0g4CDEAFkrpFbr-zNMnle_Hou90_wuf7Vdo4/edit?gid=1607116093#gid=1607116093")

fb <- gsheets2tbl(url_fb)

url_ds <- paste0("https://docs.google.com/spreadsheets/d/"
  , "1510f0Kj0g4CDEAFkrpFbr-zNMnle_Hou90_wuf7Vdo4/edit?gid=1278145622#gid=1278145622")

ds <- gsheets2tbl(url_ds)

fb <- ds %>% tarpuy_design()

url_trt <- paste0("https://docs.google.com/spreadsheets/d/"
  , "1510f0Kj0g4CDEAFkrpFbr-zNMnle_Hou90_wuf7Vdo4/edit?gid=1665653985#gid=1665653985")

traits <- gsheets2tbl(url_trt)

fbapp <- tarpuy_traits(fb, last_factor = "cols", traits)

dsg <- fbapp[[1]]

## End(Not run)
```

---

web\_table

*HTML tables for markdown documents*

---

## Description

Export tables with download, pasta and copy buttons

## Usage

```
web_table(
  data,
  caption = NULL,
  digits = 2,
  rnames = FALSE,
  buttons = NULL,
  file_name = "file",
  scrolly = NULL,
  columnwidth = "200px",
  width = "100%"
)
```

**Arguments**

data	Dataset.
caption	Title for the table.
digits	Digits number in the table exported.
rnames	Row names.
buttons	Buttons: "excel", "copy" or "none". Default c("excel", "copy")
file_name	Excel file name
scrolly	Windows height to show the table. Default "45vh"
columnwidth	Column width. Default '200px'
width	Width in pixels or percentage (Defaults to automatic sizing)

**Value**

table in markdown format for html documents

**Examples**

```
## Not run:

library(inti)

met %>%
  web_table(caption = "Web table")

## End(Not run)
```

---

yupana

*Interactive data analysis*

---

**Description**

Invoke RStudio addin to analyze and graph experimental design data

**Usage**

```
yupana(dependencies = FALSE)
```

**Arguments**

dependencies    Install package dependencies for run the app

**Details**

Yupana: data analysis and graphics for experimental designs.

**Value**

Shiny app

**Examples**

```
if(interactive()){
  inti::yupana()
}
```

---

yupana_analysis	<i>Fieldbook analysis report</i>
-----------------	----------------------------------

---

**Description**

Function to create a complete report of the fieldbook

**Usage**

```
yupana_analysis(
  data,
  last_factor = NULL,
  response,
  model_factors,
  comparison,
  test_comp = "SNK",
  sig_level = 0.05,
  plot_dist = "boxplot",
  plot_diag = FALSE,
  digits = 2
)
```

**Arguments**

data	Field book data.
last_factor	The last factor in your fieldbook.
response	Response variable.
model_factors	Model used for the experimental design.
comparison	Factors to compare
test_comp	Comprasion test c("SNK", "TUKEY", "DUNCAN")
sig_level	Significal test (default: p = 0.005)
plot_dist	Plot data distribution (default = "boxplot")
plot_diag	Diagnostic plots for model (default = FALSE).
digits	Digits number in the table exported.

**Value**

list

**Examples**

```
## Not run:  
  
library(inti)  
  
fb <- potato  
  
rsl <- yupana_analysis(data = fb  
                        , last_factor = "bloque"  
                        , response = "spad_83"  
                        , model_factors = "geno * treat"  
                        , comparison = c("geno", "treat")  
                        )  
  
## End(Not run)
```

---

yupana\_export

*Graph options to export*

---

**Description**

Function to export the graph options and model parameters

**Usage**

```
yupana_export(  
  data,  
  type = NA,  
  xlab = NA,  
  ylab = NA,  
  glab = NA,  
  ylimits = NA,  
  xrotation = c(0, 0.5, 0.5),  
  xtext = NA,  
  gtext = NA,  
  legend = "top",  
  sig = NA,  
  error = NA,  
  color = TRUE,  
  opt = NA,  
  dimension = c(20, 10, 100)  
)
```

**Arguments**

data	Result from yupana_analysis or yupana_import.
type	Plot type
xlab	Title for the axis x
ylab	Title for the axis y
glab	Title for the legend
ylimits	limits of the y axis
xrotation	Rotation in x axis c(angle, h, v)
xtext	Text labels in x axis
gtext	Text labels in group
legend	the position of legends ("none", "left", "right", "bottom", "top", or two-element numeric vector)
sig	Column with the significance
error	Show the error bar ("ste" or "std").
color	colored figure (TRUE), otherwise black & white (FALSE)
opt	Add news layer to the plot
dimension	Dimension of graphs

**Value**

data frame

**Examples**

```
## Not run:

library(inti)
library(gsheet)

url <- paste0("https://docs.google.com/spreadsheets/d/"
             , "15r7ZwcZZHbEglt1F6gSFvCTFA-CFzVBWwg3mF1RyKPs/edit#gid=172957346")
# browseURL(url)

fb <- gsheets2tbl(url)

smr <- yupana_analysis(data = fb
                      , last_factor = "bloque"
                      , response = "spad_83"
                      , model_factors = "block + geno*riego"
                      , comparison = c("geno", "riego")
                      )

gtab <- yupana_export(smr, type = "line", ylimits = c(0, 100, 2))

#> import
```

```
url <- paste0("https://docs.google.com/spreadsheets/d/"
             , "15r7ZwcZZHbEglt1F6gSFvCTFA-CFzVBWwg3mFlRyKPs/edit#gid=1202800640")
# browseURL(url)

fb <- gsheets2tbl(url)

info <- yupana_import(fb)

etab <- yupana_export(info)

info2 <- yupana_import(etab)

etab2 <- yupana_export(info2)

## End(Not run)
```

---

yupana_import	<i>Import information from data summary</i>
---------------	---

---

### Description

Graph summary data

### Usage

```
yupana_import(data)
```

### Arguments

data                    Summary information with options

### Value

list

### Examples

```
## Not run:

library(inti)
library(gsheets)

url <- paste0("https://docs.google.com/spreadsheets/d/"
             , "15r7ZwcZZHbEglt1F6gSFvCTFA-CFzVBWwg3mFlRyKPs/edit?gid=2137596914#gid=2137596914")
# browseURL(url)

fb <- gsheets2tbl(url)
```

```
info <- yupana_import(fb)

## End(Not run)
```

---

yupana\_mvr

*Multivariate Analysis*

---

## Description

Multivariate analysis for PCA and HCPC

## Usage

```
yupana_mvr(  
  data,  
  last_factor = NULL,  
  summary_by = NULL,  
  groups = NULL,  
  variables = NULL  
)
```

## Arguments

data	Field book data.
last_factor	The last factor in your fieldbook [string: NULL].
summary_by	Variables for group the analysis.
groups	Groups for color in PCA.
variables	Variables to be use in the analysis [string: NULL].

## Details

Compute and plot information for multivariate analysis (PCA, HCPC and correlation).

## Value

result and plots

## Examples

```
## Not run:  
  
library(inti)  
  
fb <- inti::potato
```



```

mv <- yupana_mvr(data = fb
  , last_factor = "geno"
  , summary_by = c("geno", "treat")
  , groups = "treat"
  , variables = c("all")
  #, variables = c("wue", "twue")
  )

mv$plot[1]

mv$data

## End(Not run)

```

---

yupana\_reshape      *Fieldbook reshape*

---

## Description

Function to reshape fieldbook according a separation character

## Usage

```

yupana_reshape(
  data,
  last_factor,
  sep,
  new_colname,
  from_var = NULL,
  to_var = NULL,
  exc_factors = NULL
)

```

## Arguments

data	Field book raw data.
last_factor	The last factor in your field book.
sep	Character that separates the last value.
new_colname	The new name for the column created.
from_var	The first variable in case you want to exclude several. variables.
to_var	The last variable in case you want to exclude several variables.
exc_factors	Factor to exclude during the reshape.

**Details**

If you variable name is `variable_evaluation_rep`. The reshape function will help to create the column `rep` and the new variable name will be `variable_evaluation`.

**Value**

data frame

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