

Package ‘tRNAscanImport’

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Title Importing a tRNAscan-SE result file as GRanges object

Version 1.31.0

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Description The package imports the result of tRNAscan-SE as a GRanges object.

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Encoding UTF-8

LazyData false

Depends R (>= 3.5), GenomicRanges, tRNA

Imports methods, stringr, BiocGenerics, Biostrings, Structstrings,
S4Vectors, IRanges, XVector, Seqinfo, rtracklayer, BSgenome,
Rsamtools

Collate 'tRNAscanImport.R' 'AllGenerics.R' 'tRNAscanImport-checks.R'
'tRNAscanImport-genome.R' 'tRNAscanImport-import.R' 'utils.R'

Suggests BiocStyle, knitr, rmarkdown, testthat, ggplot2,
BSgenome.Scerevisiae.UCSC.sacCer3

RoxygenNote 7.1.1

VignetteBuilder knitr

biocViews Software, DataImport, WorkflowStep, Preprocessing,
Visualization

BugReports <https://github.com/FelixErnst/tRNAscanImport/issues>

URL <https://github.com/FelixErnst/tRNAscanImport>

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get.tRNAPrecursor	<i>Get tRNA precursor sequences</i>
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Description

get.tRNAPrecursor retrieves tRNA precursor sequences from genomic sequences. The length of 5'- and 3'-overhangs can be specifid individually. The output is checked for validity against the tRNA sequences as reported by tRNAscan.

The chromosomes names of tRNAscan input and genome sequences must be compatible.

Usage

```
get.tRNAPrecursor(  
  input,  
  genome,  
  add.5prime = 50L,  
  add.3prime = add.5prime,  
  trim.intron = FALSE  
)
```

Arguments

- | | |
|------------------------|---|
| input | a compatible GRanges object |
| genome | a BSgenome object, a DNASTringSet object, a FaFile object or a character vector with a single value referncing a file, which can be coerced to a FaFile object. |
| add.5prime, add.3prime | the length of overhangs as a single integer value each. (default add.5prime = 50L) |
| trim.intron | TRUE or FALSE: Should intron sequences be included in the precursor sequences? (default trim.intron = FALSE) |

Value

a [DNASTringSet](#) object, containing the precursor sequences.

Examples

```
library(BSgenome.Scerevisiae.UCSC.sacCer3)
file <- system.file("extdata",
                    file = "yeast.tRNAscan",
                    package = "tRNAscanImport")
gr <- tRNAscanImport::import.tRNAscanAsGRanges(file)
genome <- getSeq(BSgenome.Scerevisiae.UCSC.sacCer3)
names(genome) <- c(names(genome)[-17], "chrmt")
get.tRNAPrecursor(gr, genome)
# this produces an error since the seqnames do not match
## Not run:
genome <- BSgenome.Scerevisiae.UCSC.sacCer3
names(genome) <- c(names(genome)[-17], "chrmt")
get.tRNAPrecursor(gr, genome)

## End(Not run)
# ... but it can also be fixed
genome <- BSgenome.Scerevisiae.UCSC.sacCer3
seqnames(genome) <- c(seqnames(genome)[-17], "chrmt")
get.tRNAPrecursor(gr, genome)
```

```
import.tRNAscanAsGRanges
```

Importing a tRNAscan output file as a GRanges object

Description

The function `import.tRNAscanAsGRanges` will import a tRNAscan-SE output file and return the information as a GRanges object. The reported intron sequences are spliced from the result by default, but can also be returned as imported.

The function `tRNAscan2GFF` formats the output of `import.tRNAscanAsGRanges` to be GFF3 compliant.

`tRNAscanID` generates a unique tRNA ID, which is like the format used in the SGD annotation

`t*AminoAcidSingleLetter*(Anticodon)*ChromosomeIdentifier**optionalNumberIfOnTheSameChromosome*`

Example: `tP(UGG)L` or `tE(UUC)E1`.

Usage

```
import.tRNAscanAsGRanges(input, as.GFF3 = FALSE, trim.intron = TRUE)
```

```
tRNAscan2GFF(input)
```

```
tRNAscanID(input)
```

Arguments

- `input`
 - `import.tRNAscanAsGRanges`: a tRNAscan-SE input file
 - `tRNAscan2GFF`: a compatible GRanges object such as the output of `import.tRNAscanAsGRanges`
- `as.GFF3` optional logical for `import.tRNAscanAsGRanges`: returns a gff3 compatible GRanges object directly. (default: `as.GFF3 = FALSE`)
- `trim.intron` optional logical for `import.tRNAscanAsGRanges`: remove intron sequences. This changes the tRNA length reported. To retrieve the original length fo the tRNA gene, use the `width()` function on the GRanges object. (default: `trim.intron = TRUE`)

Value

a GRanges object

References

Chan, Patricia P., and Todd M. Lowe. 2016. "GtRNAdb 2.0: An Expanded Database of Transfer Rna Genes Identified in Complete and Draft Genomes." *Nucleic Acids Research* 44 (D1): D184–9. doi:10.1093/nar/gkv1309.

Lowe, T. M., and S. R. Eddy. 1997. "tRNAscan-Se: A Program for Improved Detection of Transfer Rna Genes in Genomic Sequence." *Nucleic Acids Research* 25 (5): 955–64.

Examples

```
gr <- import.tRNAscanAsGRanges(system.file("extdata",
                                           file = "yeast.tRNAscan",
                                           package = "tRNAscanImport"))

gff <- tRNAscan2GFF(gr)
identical(gff,import.tRNAscanAsGRanges(system.file("extdata",
                                                    file = "yeast.tRNAscan",
                                                    package = "tRNAscanImport"),
                                           as.GFF3 = TRUE))
```

istRNAscanGRanges	<i>tRNAscan compatibility check</i>
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Description

`istRNAscanGRanges` checks whether a GRanges object contains the information expected for a tRNAscan result.

Usage

```
istRNAscanGRanges(gr)

## S4 method for signature 'GRanges'
istRNAscanGRanges(gr)
```

Arguments

`gr` the GRanges object to test

Value

a logical value

Examples

```
file <- system.file("extdata",
                    file = "yeast.tRNAscan",
                    package = "tRNAscanImport")
gr <- tRNAscanImport::import.tRNAscanAsGRanges(file)
istRNAscanGRanges(gr)
```

tRNAscanImport

tRNAscanImport: Importing tRNAscan-SE output as GRanges

Description

tRNAscan-SE can be used for prediction of tRNA genes in whole genomes based on sequence context and calculated structural features. Many tRNA annotations in genomes contain or are based on information generated by tRNAscan-SE, for example the current SGD reference genome sacCer3 for *Saccharomyces cerevisiae*. However, not all available information from tRNAscan-SE end up in the genome annotation. Among these are for example structural information, additional scores and the information, whether the conserved CCA-end is encoded in the genomic DNA. To work with this complete set of information, the tRNAscan-SE output can be parsed into a more accessible GRanges object using ‘tRNAscanImport’.

Manual

Please refer to the tRNAscanImport vignette for an example how to work and use the package: [tRNAscanImport](#)

Author(s)

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References

Chan, Patricia P., and Todd M. Lowe. 2016. “GtRNAdb 2.0: An Expanded Database of Transfer Rna Genes Identified in Complete and Draft Genomes.” *Nucleic Acids Research* 44 (D1): D184–189.. doi:10.1093/nar/gkv1309.

Lowe, T. M., and S. R. Eddy. 1997. “tRNAscan-Se: A Program for Improved Detection of Transfer Rna Genes in Genomic Sequence.” *Nucleic Acids Research* 25 (5): 955–964.

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