

# Package ‘gVenn’

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**Title** Proportional Venn and UpSet Diagrams for Gene Sets and Genomic Regions

**Version** 0.99.1

**Description** Tools to compute and visualize overlaps between gene sets or genomic regions. Venn diagrams with proportional areas are provided, while UpSet plots are recommended for larger numbers of sets. The package supports GRanges and GRangesList inputs, and integrates with analysis workflows for ChIP-seq, ATAC-seq, and other genomic interval data. It generates clean, interpretable, and publication-ready figures.

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**Encoding** UTF-8

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**RoxygenNote** 7.3.2

**URL** <https://github.com/ckntav/gVenn>

**BugReports** <https://github.com/ckntav/gVenn/issues>

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## Contents

a549_chipseq_peaks . . . . .	2
computeOverlaps . . . . .	3
exportOverlaps . . . . .	4
extractOverlaps . . . . .	5
gene_list . . . . .	6
plotUpSet . . . . .	7
plotVenn . . . . .	8
saveViz . . . . .	9
today . . . . .	10

<b>Index</b>	<b>11</b>
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a549_chipseq_peaks	<i>A549 ChIP-seq Consensus Peak Subsets (Dex, chr7)</i>
--------------------	---

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### Description

Example consensus peak subsets for MED1, BRD4, and GR after dexamethasone treatment in A549 cells. Each set has been restricted to peaks on chr7 to keep the dataset small and suitable for examples and tests.

### Usage

```
a549_chipseq_peaks
```

### Format

A GRangesList with 3 named elements:

**MED1\_Dex\_chr7** Consensus MED1 peaks (chr7 subset).

**BRD4\_Dex\_chr7** Consensus BRD4 peaks (chr7 subset).

**GR\_Dex\_chr7** Consensus GR peaks (chr7 subset).

### Details

The original full consensus peak sets are available as gzipped BED files in inst/extdata/:

- A549\_MED1\_Dex.stdchr.bed.gz
- A549\_BRD4\_Dex.stdchr.bed.gz
- A549\_GR\_Dex.stdchr.bed.gz

These are not trimmed, but for package efficiency the dataset here (a549\_chipseq\_peaks) only includes the chr7 subsets.

### Source

Internal consensus peak sets processed in A549 cells after dexamethasone stimulation.

## References

Tav C, Fournier É, Fournier M, Khadangi F, Baguette A, Côté MC, Silveira MAD, Bérubé-Simard F-A, Bourque G, Droit A, Bilodeau S (2023). "Glucocorticoid stimulation induces regionalized gene responses within topologically associating domains." *Frontiers in Genetics*. doi:10.3389/fgene.2023.1237092

## Examples

```
# Load dataset
data(a549_chipseq_peaks)
a549_chipseq_peaks

# Compute overlaps and plot
ov <- computeOverlaps(a549_chipseq_peaks)
plotVenn(ov)
```

---

computeOverlaps

*Compute Overlaps Between Multiple Sets or Genomic Regions*

---

## Description

computeOverlaps() is the main entry point for overlap analysis. It accepts either genomic region objects (GRanges/GRangesList) or ordinary sets (character/numeric vectors) and computes a binary overlap matrix describing the presence or absence of each element across sets.

## Usage

```
computeOverlaps(x)
```

## Arguments

x                    Input sets. One of:

- A GRangesList object.
- A named list of GRanges objects.
- A named list of atomic vectors (character, numeric, factor, etc.), all of the same type.

## Details

- When provided with genomic regions, the function merges all intervals into a non-redundant set (reduce()), then determines which original sets each region overlaps.
- When provided with ordinary sets (e.g., gene symbols), it collects all unique elements and records which sets contain them.

The resulting object encodes both the overlap matrix and compact category labels (e.g., "110") representing the overlap pattern of each element. These results can be directly passed to visualization functions such as plotVenn() or plotUpSet().

Internally, computeOverlaps() dispatches to either computeGenomicOverlaps() (for genomic inputs) or computeSetOverlaps() (for ordinary sets). Users are encouraged to call only computeOverlaps().

**Value**

An S3 object encoding the overlap result whose class depends on the input type:

**GenomicOverlapResult** Returned when the input is genomic (GRangesList or list of GRanges).

A list with:

- `reduced_regions`: A GRanges object containing the merged (non-redundant) intervals. Each region is annotated with an `intersect_category` column.
- `overlap_matrix`: A logical matrix indicating whether each reduced region overlaps each input set (rows = regions, columns = sets).

**SetOverlapResult** Returned when the input is a list of atomic vectors. A list with:

- `unique_elements`: Character vector of all unique elements across the sets.
- `overlap_matrix`: A logical matrix indicating whether each element is present in each set (rows = elements, columns = sets).
- `intersect_category`: Character vector of category codes (e.g., "110") for each element.

**See Also**

[plotVenn](#), [plotUpSet](#), [GRangesList](#), [reduce](#)

**Examples**

```
# Example with gene sets (built-in dataset)
data(gene_list)
ov_sets <- computeOverlaps(gene_list)
head(ov_sets$overlap_matrix)
plotVenn(ov_sets)

# Example with genomic regions (built-in dataset)
data(a549_chipseq_peaks)
ov_gr <- computeOverlaps(a549_chipseq_peaks)
head(ov_gr$overlap_matrix)
plotVenn(ov_gr)
```

---

exportOverlaps

*Export Overlap Groups to Excel*

---

**Description**

This function exports the output of `extractOverlaps()` to an Excel file, creating one sheet per overlap group. Genomic overlaps (GRanges) are converted to data frames before export.

**Usage**

```
exportOverlaps(
  grouped,
  output_dir = ".",
  output_file = "overlap_groups",
  with_date = TRUE,
  verbose = TRUE
)
```

**Arguments**

grouped	Overlap groups from extractOverlaps().
output_dir	A string specifying the output directory. Defaults to ".".
output_file	A string specifying the base filename (without extension). Defaults to "overlap_groups".
with_date	Logical (default TRUE). Whether to prepend the current date (from today) to the filename.
verbose	Logical. If TRUE, print a message with the saved path. Default TRUE.

**Value**

Overlap groups are saved to a Excel file on disk. Invisibly returns the full path to the saved file.

**Examples**

```
res <- computeOverlaps(list(A = letters[1:3], B = letters[2:4]))
grouped <- extractOverlaps(res)
exportOverlaps(grouped, output_dir = tempdir(), output_file = "overlap_groups.xlsx")
```

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extractOverlaps	<i>Extract Overlap Groups from Genomic or Set Overlap Results</i>
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---

**Description**

This function extracts subsets of intersecting elements grouped by their overlap category (e.g., "110"). For genomic overlaps, it returns a GRangesList; for set overlaps, it returns a named list of character vectors.

**Usage**

```
extractOverlaps(overlap_object)
```

**Arguments**

overlap\_object A GenomicOverlapsResult or SetOverlapsResult object.

**Value**

A named list of grouped intersecting elements:

- If input is a GenomicOverlapsResult, a GRangesList split by intersect\_category.
- If input is a SetOverlapsResult, a named list of character vectors grouped by intersect\_category.

## Examples

```
# Example with gene sets (built-in dataset)
data(gene_list)
res_sets <- computeOverlaps(gene_list)
group_gene <- extractOverlaps(res_sets)
group_gene

# Example with genomic regions (built-in dataset)
data(a549_chipseq_peaks)
res_genomic <- computeOverlaps(a549_chipseq_peaks)
group_genomic <- extractOverlaps(res_genomic)
group_genomic
```

---

gene\_list

*Example Gene Lists with Overlaps*

---

## Description

A synthetic dataset of three gene lists, created from the first 250 human gene symbols in **org.Hs.eg.db**.

## Usage

```
gene_list
```

## Format

A named list of length 3. Each element is a character vector of gene symbols:

**random\_genes\_A** 125 gene symbols.

**random\_genes\_B** 115 gene symbols.

**random\_genes\_C** 70 gene symbols.

## Source

Generated from **org.Hs.eg.db** (keys of type SYMBOL), using a reproducible random seed.

## Examples

```
data(gene_list)

# Inspect the list
str(gene_list)

# Compute overlaps and plot
ov <- computeOverlaps(gene_list)
plotVenn(ov)
```

---

`plotUpSet`*Plot an UpSet Diagram from Genomic or Set Overlap Results*

---

## Description

This function creates an UpSet plot using the ComplexHeatmap package to visualize intersections across multiple sets. Supports both GenomicOverlapsResult and SetOverlapsResult objects.

## Usage

```
plotUpSet(overlap_object, customSetOrder = NULL)
```

## Arguments

`overlap_object` A GenomicOverlapsResult or SetOverlapsResult object returned by `computeOverlaps`.

`customSetOrder` Optional. A vector specifying the order of sets to display on the UpSet diagram. The vector should contain either numeric indices (corresponding to the sets in the overlap object) or character names (matching the set names). If NULL (default), sets are displayed in decreasing order of their size (`set_size()`).

## Value

An UpSet plot object generated by `ComplexHeatmap::UpSet`.

## Examples

```
# Example with gene sets (built-in dataset)
data(gene_list)
res_sets <- computeOverlaps(gene_list)

# Default order (sets sorted by size)
plotUpSet(res_sets)

# Custom order by names
plotUpSet(res_sets, customSetOrder = c("random_genes_C",
                                       "random_genes_A",
                                       "random_genes_B"))

# Example with genomic regions (built-in dataset)
data(a549_chipseq_peaks)
res_genomic <- computeOverlaps(a549_chipseq_peaks)
plotUpSet(res_genomic)
```

plotVenn

*Plot a Venn Diagram from Genomic or Set Overlap Results***Description**

This function creates a Venn diagram using the `eulerr` package to visualize intersections across multiple sets (up to ~5). Supports both `GenomicOverlapsResult` and `SetOverlapsResult` objects.

**Usage**

```
plotVenn(
  overlap_object,
  labels = FALSE,
  legend = "right",
  fill = c("#2B70AB", "#FFB027", "#3EA742", "#CD3301", "#9370DB", "#008B8B", "#D87093"),
  quantities = list(type = "counts"),
  ...
)
```

**Arguments**

`overlap_object` A `GenomicOverlapsResult` or `SetOverlapsResult` object returned by [computeOverlaps](#).

`labels` Logical. Whether to show set labels on the diagram. Default is `FALSE`.

`legend` Position of the legend ("right", "top", "bottom", etc.) or `FALSE` to disable.

`fill` A character vector of fill colors for the sets.

`quantities` How to display intersection quantities (passed to [plot.euler](#)). Can be:

- **logical**: `TRUE` shows default counts, `FALSE` hides them.
- **vector**: custom text labels (defaults to original values from `euler()`).
- **list**: advanced options (as in `grid::grid.text()`). Use `type` to control format, combining "counts" and/or "percent" (e.g., `c("counts", "percent")`) prints counts followed by percentages in brackets). Default: `list(type = "counts")`.

... Additional arguments passed to [plot.euler](#).

**Value**

A Venn diagram plot generated by `eulerr`.

**Examples**

```
# Example with gene sets (built-in dataset)
data(gene_list)
res_sets <- computeOverlaps(gene_list)
plotVenn(res_sets)

# Example with genomic regions (built-in dataset)
data(a549_chipseq_peaks)
res_genomic <- computeOverlaps(a549_chipseq_peaks)
plotVenn(res_genomic)
```



---

`saveViz`*Save a Visualization to File (PDF, PNG, or SVG)*

---

## Description

This function saves a visualization object to a file in the specified format and directory. It supports visualizations generated by `plotVenn()`, `plotUpSet()`, `ggplot2`, or any other plot object that can be rendered using `print()` inside a graphics device. Optionally, the current date (stored in the `today` variable) can be prepended to the filename.

## Usage

```
saveViz(  
  viz,  
  output_dir = ".",  
  output_file = "figure_gVenn",  
  format = "pdf",  
  with_date = TRUE,  
  width = 5,  
  height = 5,  
  resolution = 300,  
  verbose = TRUE  
)
```

## Arguments

<code>viz</code>	A visualization object typically created by either <code>plotVenn()</code> or <code>plotUpSet()</code> , but can also be a <code>ggplot2</code> plot or any other plot object printable with <code>print()</code> .
<code>output_dir</code>	A string specifying the output directory. Defaults to ".".
<code>output_file</code>	A string specifying the base filename (without extension). Defaults to "viz_genomicVenn".
<code>format</code>	Output format. One of "pdf", "png", or "svg". Defaults to "pdf".
<code>with_date</code>	Logical (default TRUE). Whether to prepend the current date (from today) to the filename.
<code>width</code>	Width of the output file in inches. Default is 5.
<code>height</code>	Height of the output file in inches. Default is 5.
<code>resolution</code>	Resolution in DPI (only used for PNG). Default is 300.
<code>verbose</code>	Logical. If TRUE, print a message with the saved path. Default TRUE.

## Value

The visualization is saved to a file on disk. Invisibly returns the full path to the saved file.

## Examples

```
# Example with a built-in set dataset  
data(gene_list)  
ov_sets <- computeOverlaps(gene_list)  
venn_plot <- plotVenn(ov_sets)  
saveViz(venn_plot, output_dir = tempdir(), output_file = "venn_sets")
```

```
# Example with a built-in genomic dataset
data(a549_chipseq_peaks)
ov_genomic <- computeOverlaps(a549_chipseq_peaks)
upset_plot <- plotUpSet(ov_genomic)
saveViz(upset_plot, output_dir = tempdir(), output_file = "upset_genomic")

# Save as PNG instead of PDF
saveViz(upset_plot, format = "png", output_dir = tempdir(), output_file = "upset_example")

# Save as SVG
saveViz(venn_plot, format = "svg", output_dir = tempdir(), output_file = "venn_example")
```

---

today

*Today's Date at Package Load Time*

---

## Description

This variable stores the current date (in "yyyymmdd" format) at the time the package is loaded. It is useful for reproducible filenames (e.g., in `saveViz()`), and is automatically set when the package is attached.

## Usage

today

## Format

A character string (e.g., "20250624").

## Examples

```
# Print the date stored at package load
library(gVenn)
today

# Use it in a filename
paste0("venn_plot_", today, ".pdf")
```

# Index

## \* datasets

a549\_chipseq\_peaks, 2

gene\_list, 6

today, 10

a549\_chipseq\_peaks, 2

computeOverlaps, 3, 7, 8

exportOverlaps, 4

extractOverlaps, 5

gene\_list, 6

GRangesList, 4

plot.euler, 8

plotUpSet, 4, 7

plotVenn, 4, 8

reduce, 4

saveViz, 9

today, 10