

# Package ‘IdeoViz’

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**Type** Package

**Title** Plots data (continuous/discrete) along chromosomal ideogram

**Version** 1.2.0

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**Depends** Biobase, IRanges, GenomicRanges, RColorBrewer,  
rtracklayer,graphics,GenomeInfoDb

**biocViews** Visualization,Microarray

**Description** Plots data associated with arbitrary genomic intervals  
along chromosomal ideogram.

**License** GPL-2

**NeedsCompilation** no

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**Description**

Plotting discrete or continuous dataseries in the context of chromosomal location has several useful applications in genomic analysis. Examples of possible metrics include RNA expression levels, densities of epigenetic marks or genomic variation, while applications could range from the analysis of a single variable in a single context, to multiple measurements in several biological contexts (e.g. age/sex/tissue/disease context). Visualization of metrics against the chromosome could identify:

1. could identify distinctive spatial distribution that could further hypotheses about the functional role of the metric (e.g. telocentric or pericentromeric enrichment)
2. could highlight distribution differences between different groups of samples, suggesting different regulatory mechanisms; in extreme cases, visualization may identify large genomic foci of differences
3. could confirm that a quantitative difference measured between groups of interest is consistent throughout the genome (i.e. that there are no foci, and that the change is global).

This package provides a method to plot one or several dataseries against the chromosomal ideogram. It provides some simple options (vertical/horizontal orientation, display in bars or linegraphs). Data are expected to be binned; IdeoViz provides a function for user-specified bin widths. Ideograms for the genome of choice can also be automatically downloaded from UCSC using the `getIdeo()` function.

**Details**

Package:	IdeoViz
Type:	Package
Title:	Plots data (continuous/discrete) along chromosomal ideogram
Version:	0.99.1
Date:	2013-06-26
Author:	Shraddha Pai <Shraddha.Pai@camh.ca>, Jingliang Ren
Maintainer:	Shraddha Pai <Shraddha.Pai@camh.ca>
Depends:	Biobase, IRanges, GenomicRanges, RColorBrewer, rtracklayer
biocViews:	Visualization, Microarray
License:	GPL-2

**Author(s)**

Shraddha Pai <Shraddha.Pai@camh.ca>, Jingliang Ren

---

 avgByBin

*Aggregates data by genomic bins*


---

### Description

Computed mean value of binned data. This function assumes that all elements in featureData have identical width. If provided with elements of disparate widths, the respective widths are not weighted averaging. This behaviour may change in future versions of IdeoViz.

### Usage

```
avgByBin(xpr, featureData, target_GR, justReturnBins = FALSE,
         getBinCountOnly = FALSE, FUN = mean, doSampleCor = FALSE,
         verbose = FALSE)
```

### Arguments

xpr	(data.frame or matrix) Locus-wise values. Rows correspond to genomic intervals (probes, genes, etc.) while columns correspond to individual samples
featureData	(data.frame or GRanges) Locus coordinates. Row order must match xpr. Column order should be: 1. chrom, 2. locus start, 3. locus end. All elements are assumed to be of identical width. Coordinates must be zero-based or one-based, but not half-open. Coordinate system must match that of target_GR.
target_GR	(GRanges) Target intervals, with coordinate system matching that of featureData.
justReturnBins	(logical) when TRUE, returns the coordinates of the bin to which each row belongs. Does not aggregate data in any way. This output can be used as input for more complex functions with data from each bin.
getBinCountOnly	(logical) when TRUE, does not aggregate or expect xpr. Only returns number of overlapping subject ranges per bin. Speeds up computation.
FUN	(function) function to aggregate data in bin
doSampleCor	(logical) set to TRUE to compute mean pairwise sample correlation (Pearson correlation) for each bin; when TRUE, this function overrides FUN.
verbose	(logical) print status messages

### Details

This function allows the user to bin data if this hasn't already been done, and is a step involved in preparing the data for plotOnIdeo(). This function computes binned within-sample average of probes overlapping the same range. Where a range overlaps multiple bins, it gets counted in all.

**Value**

(GRanges) Binned data or binning statistics; information returned for non-empty bins only. The default for this function is to return binned data; alternately, if `justReturnBins=TRUE` or `getBinCountOnly=TRUE` the function will return statistics on bin counts. The latter may be useful to plot spatial density of the input metric.

The flags and output types are presented in order of evaluation precedence:

1. If `getBinCountOnly=TRUE`, returns a list with a single entry: *bin\_ID*: (data.frame) bin information: chrom, start, end, width, strand, index, and count. "index" is the row number of *target\_GR* to which this bin corresponds
2. If `justReturnBins=TRUE` and `getBinCountOnly=FALSE`, returns a list with three entries:
  - (a) *bin\_ID*: same as *bin\_ID* in output 1 above
  - (b) *xpr*:(data.frame) *B*-by-*n* columns where *B* is total number of [*target\_GR*,*featureData*] overlaps (see next entry, *binmap\_idx*) and *n* is number of columns in *xpr*; column order matches *xpr*. Contains sample-wise data "flattened" so that each [*target*,*subject*] pair is presented. More formally, entry [*i*,*j*] contains expression for overlap of row *i* from *binmap\_idx* for sample *j* (where  $1 \leq i \leq B$ ,  $1 \leq j \leq n$ )
  - (c) *binmap\_idx*:(matrix) two-column matrix: 1) *target\_GR* row, 2) row of *featureData* which overlaps with index in column 1. (matrix output of `GenomicRanges::findOverlaps()`)
3. Default: If `justReturnBins=FALSE` and `getBinCountOnly=FALSE`, returns a GRanges object. Results are contained in the `elementMetadata` slot. For a dataset with *n* samples, the table would have (*n*+1) columns; the first column is *bin\_count*, and indicates number of units contained in that bin. Columns (2:(*n*+1)) contain binned values for each sample in column order corresponding to that of *xpr*.  
For `doSampleCor=TRUE`, result is in a metadata column with name "mean\_pairwise"cor". Bins with a single datapoint per sample get a value of NA.

**Author(s)**

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**See Also**

`getIdeo()`, `getBins()`

**Examples**

```
ideo_hg19 <- getIdeo("hg19")
data(GSM733664_broadPeaks)
chrom_bins <- getBins(c("chr1", "chr2", "chrX"), ideo_hg19, stepSize=5*100*1000)
# default binning
mean_peak <- avgByBin(data.frame(value=GSM733664_broadPeaks[,7]), GSM733664_broadPeaks[,1:3], chrom_bins)
# custom function
median_peak <- avgByBin(data.frame(value=GSM733664_broadPeaks[,7]), GSM733664_broadPeaks[,1:3], chrom_bins, FUN=median)
# mean pairwise sample correlation
data(binned_multiSeries)
bins2 <- getBins(c("chr1"), ideo_hg19, stepSize=5e6)
samplecor <- avgByBin(mcols(binned_multiSeries)[,1:3], binned_multiSeries, bins2, doSampleCor=TRUE)
# just get bin count
```

```
binstats <- avgByBin(data.frame(value=GSM733664_broadPeaks[,7]), GSM733664_broadPeaks[,1:3], chrom_bins, getBin
```

---

binned\_fullGenome      *Data for example 3.*

---

### Description

Simulated data spanning all autosomes and X,Y chromosomes of the human genome (build hg18). Values consist of a single dataserie of random uniform distribution between -1 and + 1. The chromosomes are tiled in 1Mb bins and coordinates are one-based.

### Usage

```
data(binned_fullGenome)
```

### Source

Simulated data, generated by Shraddha Pai

### Examples

```
data(binned_fullGenome)
head(binned_fullGenome)
seqlevels(binned_fullGenome)
```

---

binned\_multiSeries      *Data for vignette example 1.*

---

### Description

A simulated dataserie spanning chr1,chrX,chrY of the human genome (build hg18). Values consist of five series constructed to show mostly random behaviour with the exception of elevated signal in a few regions. The chromosomes are tiled in 1Mb bins and coordinates are one-based.

### Usage

```
data(binned_multiSeries)
```

### Source

Simulated data, generated by Shraddha Pai

### Examples

```
data(binned_multiSeries)
head(binned_multiSeries)
```

---

binned\_singleSeries     *Data for example 2.*

---

### Description

Simulated data spanning 3 human chromosomes and varying in a random uniform distribution between -1 and +1.

### Usage

```
data(binned_singleSeries)
```

### Source

Simulated data by Shraddha Pai

### Examples

```
data(binned_singleSeries)
head(binned_singleSeries)
```

---

getBins                     *getBins*

---

### Description

Get uniformly-sized bins of specified width

### Usage

```
getBins(chroms, ideo, binLim = NULL, stepSize)
```

### Arguments

chroms	(character) chromosomes to generate bins for
ideo	(data.frame) ideogram table as generated by <code>getIdeo()</code> . See that function for details.
binLim	(numeric, length 2) [start, end] of genomic range to generate bins for. A value of NULL results in binning of entire chromosome
stepSize	(integer) bin size in bases

### Details

This is a helper function used to generate binned data for `plotOnIdeo()`. It takes the chromosome-wide extents from *ideo*, which is essentially the *cytoBandIdeo* table from UCSC browser with the header as the first row. A use case is to generate bins using this function and supply the output to `avgByBin()` to bin the data.

**Value**

(GRanges) bin ranges in 1-base coordinates

**Author(s)**

Shraddha Pai <Shraddha.Pai@camh.ca>, Jingliang Ren

**See Also**

getIdeo(), avgByBin()

**Examples**

```
ideo_hg19 <- getIdeo("hg19")
chrom_bins <- getBins(c("chr1", "chr2", "chrX"), ideo_hg19, stepSize=5*100*1000)
```

---

getIdeo

*Download ideogram table from UCSC*

---

**Description**

Download table containing chromosomal extent and band locations from the UCSC genome browser

**Usage**

```
getIdeo(ideoSource)
```

**Arguments**

ideoSource (character) Genome build for data (e.g. mm10).

**Details**

Uses `rtracklayer` to retrieve the `cytoBandIdeo` table from the UCSC genome browser. The `cytoBandIdeo` table contains chromosomal ideogram information and is used to graph the chromosomal bands in `plotOnIdeo()`. This table is provided as input to `plotOnIdeo()`. In the case where the user bins the data, the output of this function can also be used as input to generate bin coordinates for binning the data (see `avgByBin()`).

**Value**

(data.frame) ideogram table

**Author(s)**

Shraddha Pai <Shraddha.Pai@camh.ca>, Jingliang Ren

**See Also**

```
avgByBin().getBins()
```

**Examples**

```
getIdeo("mm9")
```

---

GSM733664\_broadPeaks *Data for vignette example 4.*

---

**Description**

Broadpeaks file mapping H3K9me3 marks in human lymphoblastoid cells (peaks from chr1, chr2, and chrX).

**Usage**

```
data(GSM733664_broadPeaks)
```

**Details**

GEO accession GSM733664, subset containing chr1,chr2,and chrX peaks.

**References**

ENCODE Project Consortium, Bernstein BE, Birney E, Dunham I, Green ED, Gunter C, Snyder M. An integrated encyclopedia of DNA elements in the human genome. *Nature*. 2012 Sep 6;489(7414):57-74.

**Examples**

```
data(GSM733664_broadPeaks)
head(GSM733664_broadPeaks)
```

---

hg18\_ideo *Ideogram table for hg18*

---

**Description**

Cytoband information for all chromosomes in human genome build hg18. Used for vignette examples.

**Usage**

```
data(hg18_ideo)
```



**Source**

UCSC genome browser.

**Examples**

```
data(hg18_ideo)
head(hg18_ideo)
```

---

plotChromValuePair      *Plot a chromosome-value pair*

---

**Description**

Base function which plots the ideogram and superimposed data for a single chromosome. plotOnIdeo() calls this function and stacks the resulting output.

**Usage**

```
plotChromValuePair(chrom, cytoTable, bpLim, vertical, values_GR,
  val_range, col, value_cols = "values", default_margins, addScale,
  ablines_y, smoothVals, span=0.03, verbose = FALSE, ...)
```

**Arguments**

chrom	(character) chromosome(s) to create ideograms for
cytoTable	(data.frame) loaded ideogram table. (see <i>ideoTable</i> argument to plotOnIdeo())
bpLim	(numeric) (aka xlim); display only a section of the chromosome and the corresponding values
vertical	(logical) if TRUE, chromosomes will be plotted vertically
values_GR	(GenomicRanges) data to be plotted must be in metadata columns
val_range	(numeric) (aka ylim); y-axis scale for data series
col	(character) colour for series
value_cols	(character) column name for series to plot
default_margins	(numeric) page inner margins (in inches)
addScale	(logical) if FALSE, bp positions will be hidden
ablines_y	(numeric) when specified, will draw reference lines on the y-axis
smoothVals	(logical) when T applies loess() to each series
span	(numeric) span argument for loess function
verbose	(logical) print messages
...	arguments to axis(), line(), and rect()

**Details**

Plots one unit of chromosome ideogram with dataseries superimposed. Usually, the user can avoid this function and directly call `plotOnIdeo()`. However, this function may be used in cases where further plot customization is required.

**Author(s)**

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**See Also**

`plotOnIdeo()`

**Examples**

```
data(hg18_ideo)
data(binned_multiSeries)
layout(matrix(1:2, byrow=TRUE, ncol=1), heights=c(2.5, 1))
plotChromValuePair("chr1", hg18_ideo,
  values_GR=binned_multiSeries, value_cols=colNames(mcols(binned_multiSeries)), plotType='lines',
  col=1:5, val_range=c(0, 10), bplim=NULL, vertical=FALSE, addScale=TRUE, ablines_y=NULL,
  smoothVals=FALSE, default_margins=c(0.5, .5, .1, .1))
```

---

plotOnIdeo

*Plot data superimposed on chromosomal ideogram*

---

**Description**

Main function to plot binned data alongside chromosomal ideogram.

**Usage**

```
plotOnIdeo(chrom = stop("enter chromosome(s) to plot"), ideoTable,
  values_GR, value_cols = "values", plotType = "lines", col = "orange",
  bplim = NULL, val_range = NULL, addScale = TRUE, scaleChrom = TRUE,
  vertical = FALSE, addOnetoStart = TRUE, smoothVals = FALSE, cex.axis = 1, plot_title = NULL, ab1
```

**Arguments**

chrom	(character) chromosome(s) to create ideograms for
ideoTable	(data.frame) ideogram table. See <code>getIdeo()</code>
values_GR	(GenomicRanges) data to be plotted must be in metadata columns
value_cols	(character) which series to plot. Should be column names of the <code>mcols()</code> slot of <code>values_GR</code>
plotType	(character) Plot type for each series. Values can be "lines" or "rect" to plot lines or barplots respectively. The latter is not recommended when several series are to be plotted on the same axis.)

col	(character) vector of colors for data series
bpLim	(numeric) (xlim); display only a section of the chromosome and the corresponding values
val_range	(numeric) (ylim); y-axis scale for data series
addScale	(logical) if TRUE, bp positions will be shown along the chromosomes. This feature should be turned off if numerous chromosomes' worth of data are being plotted and all objects don't fit on the final graphics device.
scaleChrom	(logical) if FALSE, all chroms will display as the same size. scaleChrom will be ignored if bpLim is not NULL
vertical	(logical) if TRUE, chromosomes will be plotted vertically
addOneToStart	(logical) if TRUE, adds 1 to chromStart. Useful to convert data in half-open coordinates - which is all data from the UCSC genome browser, including cytoBandIdeo, into 1-base.
smoothVals	(logical) if T, smoothes each trendline. Currently hard-coded to lowess smoothing with span=0.03
cex.axis	(integer) axis font size
plot_title	(character) title for overall graph
ablines_y	(numeric) when supplied, draws reference lines on the y-axis
cex.main	(numeric) font size for plot title
...	other graphing options for barplot (i.e. main="Values", to title bar plot "Values")

## Details

plotOnIdeo() is the main function of this package. It is the one the end-user is expected to call to generate plots. Input is provided as a *GRanges* object (values\_GR), with data to be plotted contained in its metadata slot. The user is responsible for providing pre-binned data, if binning is required. Data can also be binned using the avgByBin() function in this package. The ideogram table (ideoTable) is the same as the *cytoBandIdeo* table available from the UCSC genome browser database for a given genome is a can be either automatically downloaded from UCSC (see getIdeo()) or read in from a local-file and passed to this function.

There are numerous arguments which control the appearance of the plot. The main decision points are:

1. vertical: Whether the entire plot should have a horizontal or vertical orientation
2. plotType: Whether each dataseries should be shown as a trendline or as a barplot. The latter is not recommended for cases where multiple series are to be plotted on the same axis.

Other considerations:

- The size of the graphics device limits the number of chromosomes that can be plotted. A simple solution may be to set addScale=FALSE. However, it is recommended to call plotOnIdeo() multiple times, and plotting a fewer number of chromosomes on each page.
- The code expects coordinates of values\_GR to be in 1-base. Set addOneToStart=TRUE if supplied coordinates are in 0-base.

**Author(s)**

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**Examples**

```
data(binned_multiSeries)
data(hg18_ideo)
plotOnIdeo(chrom=seqlevels(binned_multiSeries), ideoTable=hg18_ideo, values_GR=binned_multiSeries, value_cols=cc
```

---

wins

*Data for vignette example 1.*

---

**Description**

A simulated dataseries spanning three chromosomes, and containing five series. The chromosomes are tiled in 1Mb windows.

**Usage**

```
data(wins)
```

**Source**

Simulation by Shraddha Pai

**Examples**

```
data(wins)
head(wins)
```

---

wins\_discrete

*Data for example 2.*

---

**Description**

Simulated data spanning 3 human chromosomes and varying in a random uniform distribution between -1 and +1.

**Usage**

```
data(wins_discrete)
```

**Source**

Simulated data by Shraddha Pai

**Examples**

```
data(wins_discrete)
head(wins_discrete)
```

---

```
wins_entiregenome
```

*Data for example 3.*

---

**Description**

Simulated data spanning all human chromosomes. Values follow random uniform distribution between  $-1 \text{ nd} + 1$ .

**Usage**

```
data(wins_entiregenome)
```

**Source**

Simulated data, generated by Shraddha Pai

**Examples**

```
data(wins_entiregenome)
head(wins_entiregenome)
seqlevels(wins_entiregenome)
```

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